Zootaxa 2544: 1–53 (2010) www.mapress.com/zootaxa/

Copyright © 2010 · Magnolia Press

Article



Helminths of the American bullfrog, *Lithobates catesbeianus* (Shaw, 1802), from Pawnee Lake, Lancaster, Nebraska, USA with a checklist of its helminth parasites

ROSARIO MATA-LÓPEZ¹, LUIS GARCÍA-PRIETO² AND VIRGINIA LEÓN-RÈGAGNON³

¹Departamento de Biología Evolutiva, Facultad de Ciencias, Universidad Nacional Autónoma de México, CP. 04510, Mexico D.F., Mexico; e-mail: gorgoderina@yahoo.com.mx

²Laboratorio de Helmintología, Instituto de Biología, Universidad Nacional Autónoma de México. AP. 70-153, C.P. 04510, México D.F., Mexico.

³ Estación de Biología Chamela, Instituto de Biología, Universidad Nacional Autónoma de México. A.P. 21, C.P. 48980, San Patricio, Jalisco, Mexico.

Table of contents

Introduction	
Materials and method	2
Results	2
Checklist of helmith parasites of <i>Lithobates catesbeianus</i> along its natural and introduced distribution range	3
Digenea	3
Monogenea	20
Cestoidea	21
Acanthocephala	23
Nematoda	24
Discussion	
Acknowledgments	39
References	39
Index	51

Abstract

The American bullfrog, *Lithobates catesbeianus*, is one of the most intensively studied host species. However, most of the records related to the parasites of this anuran are scattered through the literature. The purpose of our study is twofold: 1) to list the helmiths of this host from Pawnee Lake, Lancaster Co., Nebraska, USA, and 2) To compile all published records and those contained by several scientific collections to construct a checklist of helminth parasites associated with the American bullfrog through its range in both native and introduced geographical areas. Twenty-seven specimens of *L. catesbeianus* were collected and examined for helminth parasites; 20 frogs were infected. Nine species of helminths were collected: 4 digeneans: *Haematoloechus coloradensis*, *H. parviplexus*, *Gorgoderina attenuata*, and *Glypthelmins quieta*, and 5 nematodes: *Rhabdias ranae*, *Spinitectus gracilis*, *Cosmocercoides variabilis*, *Spiroxys* sp., and an unidentified ascarid nematode. Pawnee Lake represents a new locality record for *S. gracilis*. The digenean *H. parviplexus* had the highest prevalence and mean abundance (33.3% and 3, respectively). As a result of our study, the number of helminth taxa known for this host is 159 (75 digeneans, 4 monogeneans, 10 cestodes, 7 acanthocephalans, and 63 nematodes); these records come from 6 countries (Canada, Cuba, Japan, Korea, United Kingdom, and USA). Although well documented, the helminthological record of this host species could increase after further inventory work in poorly sampled regions.

Key words: Lithobates catesbeianus, American bullfrog, Pawnee Lake, helminths, Nematoda, Digenea, checklist

Introduction

The American bullfrog, *Lithobates catesbeianus* (Shaw, 1802) (previously *Rana catesbeiana* Shaw, 1802, see Frost *et al.* 2006), is native to North America, occurring from sea level to 2,740 m elevation, from southern Atlantic Canada to eastern Colorado and eastern New Mexico and it has been introduced to many localities in the western United States as well as in 26 countries around the world (Santos-Barrera *et al.* 2006). To the best of our knowledge, only one study compiles the helminthological record of the American bullfrog in North America, listing 95 helminth species (Andrews *et al.* 1992). For hosts from Nebraska, 22 helminth species have been recorded (Brooks 1974, 1975a, 1976, 1977; Snyder & Tkach 2001; León-Règagnon & Brooks 2003; León-Règagnon *et al.* 2005; Brooks *et al.* 2006a; Bolek & Janovy 2007). The purpose of this study is to describe helminth parasites of *L. catesbeianus* in Pawnee Lake, Lancaster County, Nebraska, as well as to summarize and update the helminth records of this host species.

Materials and method

A total of 27 adult frogs were collected by hand on September, 2001 from a permanent, anthropogenic impoundment (Pawnee Lake), Lancaster Co., Nebraska (40°51' 31.6" N, 96° 53' 20.2" W). Amphibians were collected by hand and placed in plastic containers, transported alive to the Harold W. Manter Laboratory of Parasitology (HWML), University of Nebraska, Lincoln, Nebraska and killed with an overdose of sodium pentobarbital within 48 hr of capture. The body cavity was opened and internal organs (lungs, stomach, intestine, gall bladder, liver, and urinary bladder) were removed, placed in individual Petri dishes with 0.65% saline solution and examined for helminths with a stereomicroscope. The integument and mouth cavity were also searched for helmiths. Helminths were counted in situ, collected and fixed with hot formalin 4% for morphological studies. Digeneans were stained with Mayer's paracarmin, dehydrated, cleared in a gradual series of methyl salicilate and mounted in Canada balsam. Nematodes were cleared with glycerol or Amman's lactophenol and mounted on temporary slides. Helminth infections were characterized following Bush et al. (1997). Hosts were initially fixed with 10% formalin, washed with tap water, stored in 70% ethanol and deposited in the collection of amphibians and reptiles of the Zoological Division of the University of Nebraska State Museum, with the following accession numbers: ZM 23694-23707, 23716-23728. Helminth voucher specimens were deposited in the Colección Nacional de Helmintos (CNHE), Instituto de Biología, Universidad Nacional Autónoma de México, Mexico City. To create the checklist, we used the information from a retrospective bibliographical search, using different databases (CAB Abstracts, Biological Abstracts, Zoological Record, and ISI Web of Science) and searches in data bases of parasite collections. Larval stages are indicated by asterisk (*). Data are updated to October, 2009. The following acronyms are used reffering to parasitological collections: CMNPA: Canadian Museum of Nature, Parasitological Collection, Ottawa, Canada; CZACC: Colección del Instituto de Zoología, Academía de Ciencias de Cuba, Havana, Cuba; HWML: Harold W. Manter Laboratory of Parasitology, Lincoln, Nebraska, USA; IES: Instituto de Ecología y Sistematica, Havana, Cuba; NBM: New Brunswick Museum, New Brunswick, Canada; NSMT: National Science Museum, Tokyo, Japan; USNPC: US National Parasite Collection, Maryland, USA.

Results

Twenty seven American bullfrogs were examined (5.144-23.76±25.87 gr and 34-125 (61.78±17.68) mm snout-vent). Thirteen were males, 9 females and for 5, the sex could not be recognized. Of these 20 were parasitized (74%, 44.44% harbored 0-1 species of helminths). Two hundred and two specimens distributed across 9 helminth species were recovered, 4 digenean species: *Haematoloechus coloradensis* (Cort, 1915), *H. parviplexus* (Irwin, 1929), *Gorgoderina attenuata* (Stafford, 1902), and *Glypthelmins quieta* (Stafford, 1900), and 5 nematode species: *Rhabdias ranae* Walton, 1929, *Spinitectus gracilis* Ward & Magath, 1917, *Cosmocercoides variabilis* (Harwood, 1930), *Spiroxys* sp., and an unidentified ascarid nematode. The percentage of parasitism

by organ was: lungs 49.5%, stomach 23.27%, small intestine 12.87%, mesentery 9.4%, large intestine 3.96, and urinary bladder with 0.9%. The lungs were the most parasitized organs with 3 helminth species (*H. coloradensis*, *H. parviplexus* and *R. ranae*).

Three frogs showing site specific co-infection including *R. ranae* and *H. parviplexus*, *R. ranae* and *C. variabilis*, and *H. parviplexus* and *H. colorandensis*. Adult stages were predominant in both helminth classes, nematodes had the highest prevalence values in the overall sample; however, individually *H. parviplexus* showed the highest prevalence and mean abundance (33.3% and 3, respectively).

In the present checklist, 159 helminth taxa recorded as parasites of *L. catesbeianus* are listed, including digeneans (75 taxa), followed by nematodes (63 taxa), cestodes (10 taxa), acanthocephalans (7 taxa), and finally monogeneans with 4 taxa. Most of the helminth species enlisted was recorded in adult stage (111), while 48 taxa represent larval stages. Intestine and lungs constitute the most parasitized habitats in this host species, harbored 59 and 18 helminth taxa, respectively.

Checklist of helmith parasites of L. catesbeianus along its natural and introduced distribution range

Digenea van Beneden, 1858

Alaria sp.*

Site of infection: muscle, body cavity.

Recruitment: penetration.

Distribution: Canada: Ontario (Walters *et al.* 1975, experimental). USA: Arizona (Goldberg *et al.* 1998). **Specimens in collections:** USNPC: 87070.

Remarks: Goldberg *et al.* (1998) did not consider this genus as typical of anurans, because it does not reach maturity in frogs. Frogs act as second intermediate host; definitive hosts are mammals.

Alaria arisaemoides Agustine & Uribe, 1927*

Site of infection: lungs, muscles, stomach.
Recruitment: penetration.
Distribtion: Canada: New Brunswick (McAlpine 1997; McAlpine & Burt, 1998a).
Specimens in collections: NBM.

Alaria marcianae (La Rue, 1917)*

Site of infection: muscle.
Recruitment: penetration.
Distribution: USA: Louisiana (Shoop 1985, experimental life cycle); Michigan (Bosma 1934).
No specimens in collections.
Remarks: Tadpoles have been reported as experimental second intermediate host of several species of *Alaria* (Johnson 1968).

Alaria mustelae Bosma, 1931*

Site of infection: muscle. Recruitment: penetration. Distribution: USA: Michigan (Bosma 1934, experimental life cycle).

Specimens in collections: USNPC: 8553.

Remarks: Bosma (1934) and Johnson (1979) described the life cycle of *Alaria* spp. from experimentally infected tadpoles and naturally infected frogs.

Allassostomoides chelydrae (MacCallum, 1919)

Site of infection: rectum.

Recruitment: penetration.

Distribution: USA: Nebraska (Brooks 1975b; Brooks 1976; Brooks et al. 2006a).

No specimens in collections.

Remarks: Some authors have been considered this species synonym of *A. parvus* (Stunkard 1924; Travassos 1934; Skrjabin 1947); however, Yamaguti (1958, 1971) recognized it as a valid species. *Allassostomoides chelydrae* was re-described by Brooks (1975b) based on specimens from multiple host species.

Allassostomoides parvus (Stunkard, 1916)

Site of infection: digestive system, colon, cloaca, rectum.

Recruitment: penetration.

Distribution: USA: Illinois (Beaver 1929); Louisiana (Bennett 1938); Nebraska (Brooks 1975a; Brooks 1976; Brooks *et al.* 2006a).

Specimens in collections: HWML: 20103; USNPC: 95702.

Apharyngostrigea pipientis (Faust, 1918)*

Site of infection: body cavity.

Recruitment: penetration.

Distribution: Canada: New Brunswick (McAlpine 1997).

Specimens in collections: NBM: 1410.

Remarks: This species has also been recorded in *Pseudacris triseriata* from Canada (Goldberg *et al.* 2002) and *Rana pipiens* from USA (Goldberg *et al.* 2001).

Auridistomum chelydrae (Stafford 1900)*

Site of infection: not specified.

Recruitment: penetration.

Distribution: USA: Oklahoma (Ralph 1938).

Specimens in collections: USNPC: 9260 (syntypes).

Remarks: This species was recorded as *Tetrapapillatrema concavocorpa* which is considered a synonym of *Auridistomum chelydrae* (Yamaguti 1971).

Brachycoelium salamandrae (Fröelich, 1789)

Site of infection: intestine.

Recruitment: ingestion. **Distribution:** USA: Georgia (Parker 1941).

No specimens in collections.

Remarks: Recorded as *Brachycoelium louisianae* Byrd, 1937, species synonymyzed with *B. salamandrae* by Rankin (1938) but Yamaguti (1971) included it among valid species of the genus. Reciently Yildirimhan *et al.* (2005) returned *B. louisianae* to synonymy with *B. salamandrae*.

Bunodera sp.*

Site of infection: intestine.

Recruitment: ingestion.

Distribution: USA: Kansas (USNPC).

Specimens in collections: USNPC: 91220.

Remarks: Species of this genus are typical parasites of freshwater fishes (Choudhury & León-Règagnon 2005).

Cephalogonimus sp.

Site of infection: intestine.

Recruitment: ingestion.

Distribution: USA: Kansas (USNPC).

Specimens in collections: USNPC: 91221.

Remarks: This is the only genus of the family Cephalogonimidae represented in amphibian hosts. Species of this genus are distributed from Canada to Brazil (Yamaguti 1971; Rodriguez-Ortíz *et al.* 2004; Brooks *et al.* 2006a).

Cephalogonimus americanus Stafford, 1902

Site of infection: intestine.

Recruitment: ingestion.

Distribution: USA: Massachusetts (Rankin 1945); Arizona (Goldberg *et al.* 1998); undetermined (USNPC).

Specimens in collections: USNPC: 51431.

Remarks: Adult and juvenile specimens of this species have been recorded as parasites of Ranidae and Ambystomatidae (Paredes-León *et al.* 2008). Tadpoles of *Anaxyrus fowleri* (Hinckley) were experimentally infected by Lang (1968).

Cephalogonimus brevicirrus Ingles, 1932

Site of infection: intestine.

Recruitment: ingestion. **Distribution:** USA: Nevada (Brooks, 1976). **No specimens in collections.**

Clinostomum sp.*

Site of infection: muscle, mesentery. **Recruitment:** penetration.

Distribution: Canada: Ontario (USNPC). USA: Arizona (Goldberg *et al.* 1998); Louisiana (USNPC); Michigan (Muzzall 1991; USNPC); Texas (Yoder & Gomez 2007; HWML); undetermined locality (Lemke *et al.* 2008).

Specimens in collections: HWML: 48351. USNPC: 50105, 51495, 81461, 87071.

Remarks: Freshwater fish species represent the most common intermediate hosts for species of this genus.

Clinostomum attenuatum Cort, 1913*

Site of infection: peritoneum, body cavity.
Recruitment: penetration.
Distribution: USA: Oklahoma (Trowbridge & Hefley 1934); undetermined (HWML).
Specimens in collections: HWML: 30326, 40122.
Remarks: The definitive hosts of this species are several fish-eating birds.

Clinostomum marginatum Rudolphi, 1819*

Site of infection: peritoneum.

Recruitment: penetration.

Distribution: Kansas (Jinks & Johnston 1971).

No specimens in collections.

Remarks: Baer (1933) and Ukoli (1966) synonymyzed *C. marginatum* with *C. complanatum* (Rudolphi, 1819), but recently both species were separated based on differences in ribosomal DNA (Dzikowski *et al.* 2004).

Cystagora tetracystis (Gastaldi, 1854)*

Site of infection: muscles of throat.
Recruitment: unknown.
Distribution: Canada (Stafford 1900, 1905; CMNPA).
Specimens in collections: CMNPA: 1900-1680, 1900-1681.
Remarks: Yamaguti (1971) listed this species among the larval digeneans of amphibians.

Diplodiscus sp.

Site of infection: intestine.

Recruitment: ingestion.

Distribution: USA: Indiana (USNPC).

Specimens in collections: USNPC: 51953.01.

Remarks: In accordance with Jones *et al.* (2005), species of *Diplodiscus* are distributed in Europe, Asia and Africa. Identification of these specimens needs to be confirmed.

Echinostoma trivolvis (Cort, 1914)*

Site of infection: kidney.

Recruitment: penetration.

Distribution: Canada: New Brunswick (McAlpine 1997). USA: Pennsylvania (Fried & Bradford, 1997). **Specimens in collections:** NBM: 1493.

Remarks: Identification of Canada material was based on adult specimens obtained from experimental infection in *Gallus gallus*.

Euryhelmis squamula (Rudolphi, 1819)*

Site of infection: skin.

Recruitment: penetration.

Distribution: USA: Washington (USNPC).

Specimens in collections: USNPC: 44419.

Remarks: Specific identification of these metacercariae was based on experimental infection in mice (USNPC).

Glypthelmins sp.

Site of infection: intestine.

Recruitment: ingestion.

Distribution: USA: Nebraska (HWML); Oregon (USNPC); Wisconsin (HWML).

Specimens in collections: HWML: 19720, 19722, 19723, 23832. USNPC: 66142.

Remarks: Based on morphological and molecular evidence, Razo-Mendivil and Pérez-Ponce de León (2008) redefined the genus *Glypthelmins*, including only 10 species; from these, *G. quieta*, *G. californiensis*, *G. intestinalis*, *G. hyloreus*, *G. pennsylvaniensis*, *G. parva*, *and G. shastai* are distributed in USA.

Glypthelmins californiensis (Cort, 1919)

Site of infection: intestine.
Recruitment: ingestion.
Distribution: not specified (HWML).
Specimens in collections: HWML: 31390.
Remarks: Valid species following Razo-Mendivil and Pérez-Ponce de León (2008).

Glypthelmins quieta (Stafford, 1900)

Site of infection: intestine.

Recruitment: ingestion.

Prevalence, mean intensity, and intensity range: 25.9%, 3.71, 1-13.

Specimens deposited: CNHE: 5374.

Distribution: Canada: New Brunswick (McAlpine 1997a, 1997b; 1998; McAlpine & Burt 1998a; NBM); Ontario (Stafford 1900; Stafford 1905; Walton 1938; Walton 1947); Quebec (CMNPA). Cuba: Havana (Odening 1968; Martínez *et al.* 1982). USA: Arkansas (Rosen & Manis 1976; USNPC); California (Goldberg & Bursey 2002a); Florida (Manter 1938); Georgia (USNPC; HWML; Parker 1941; Sullivan 1972; Sullivan 1976); Illinois (Miller 1930; Leigh 1937a, 1937b, 1946; Andrews *et al.* 1992; USNPC); Indiana (Lank 1971); Iowa (Ulmer 1970); Kansas (Jinks & Johnston 1971); Louisiana (Bennett 1938; Sullivan 1972; Sullivan 1976; HWML); Massachusetts (Rankin 1944a); Michigan (Najarian 1955; Muzzall 1991); Mississippi (Sullivan 1972; Sullivan 1976; Brooks 1979; HWML); Nebraska (Brooks 1975a; Brooks 1976; Brooks 1979; Brooks *et*

al. 2006a; Razo-Mendivil *et al.* 2006; HWML); Nevada (Babero & Golling 1974); New England (Rankin 1944a; Leigh & Van Cleave 1945); North Carolina (Brandt 1936; Rankin 1944a; Leigh & Van Cleave 1945); Ohio (Odlaug 1954; Ashton & Rabalais 1978); Oklahoma (Trowbridge & Hefley 1934; Brooks 1979); Oregon (O'Grady 1987; USNPC); Seattle (Rankin 1944a; Leigh & Van Cleave 1945); South Carolina (USNPC); Texas (Harwood 1932; Slagle 1966; Hollis 1972; Mayberry *et al.* 2000; Smythe & Font 2001); Virginia (Britt 1947; Campbell 1968); Washington (Rankin 1944a; Leigh & Van Cleave 1945); Winsconsin (Schell 1962); undetermined locality (Walton 1938, 1947; USNPC).

Specimens in collections: CMNPA: 1900-1696, 1900-1697, 1900-1698. HWML: 19721, 20174-20178, 20183, 20187, 20229, 20643, 20925, 20938, 20956, 22675, 31273. NBM: 746. USNPC: 66142.02, 72269, 72270, 82012, 84282, 84283, 84802, 84814, 91248, 95554.

Remarks: Type species of the genus *Glypthelmins* sensu stricto (Razo-Mendivil *et al.* 2006). Harwood (1932) described *Glypthelmins subtropica* Harwood, 1932 based on specimens from *L. catesbeianus* (USNPC: 30878); this name was used also by Parker (1941); however, in a recent phylogenetic analysis and revision of the genus, this species was considered synonym of *G. quieta* (Razo-Mendivil *et al.* 2006; Razo-Mendivil & Pérez-Ponce de León 2008).

Gorgodera sp.

Site of infection: intestine, urinary bladder.

Recruitment: ingestion.

Distribution: Canada: Ontario (CMNPA); Quebec (Fantham & Portes 1948). USA: Kansas (HWML); Louisiana (HWML); Michigan (Hunt 1952); New York (USNPC); West Virginia (NBM).

Specimens in collections: CMNPA: 1900-1617c. HWML 22682, 24896. NBM: 1376. USNPC: 36360.

Remarks: This genus is apparently distributed in the Nearctic, Palearctic and Oriental Realms (Yamaguti 1971). Some species have been found in sympatry with species of *Gorgoderina* in *L. catesbeianus* (Odlaug 1937; Andrews *et al.* 1992). Material from Michigan was obtained experimentally, and recorded as *Gorgodera vivata* Hunt, 1952, which is *nomen nudum* (Yamaguti 1971).

Gorgodera amplicava Looss, 1899a

Site of infection: kidneys and urinary bladder (in adult specimens); bladder, excretory ducts, stomach, intestine, colon, cloaca, Wolffian duct, oviduct, mesonephroi (in young specimens).

Recruitment: ingestion.

Distribution: Canada: Ontario (Bensley, 1897; Looss 1899a, 1899b; Stafford 1903; Cort 1912; CMNPA; USNPC). USA: Arkansas (Parker 1941; Rosen & Manis 1976); Illinois (Andrews *et al.* 1992; USNPC); Indiana (Lank 1971); Iowa (Ulmer 1970); Kentucky (Parker 1941); Louisiana (Bennett 1938; Goodchild 1954, 1955; HWML); Massachusetts (Odlaug 1937; Goodchild 1945, 1948, 1950, 1955); Michigan (Krull 1935); Mississippi (Brooks 1979); Missouri (Goodchild 1948,1950, 1954; HWML); Nebraska (Brooks 1976; Brooks *et al.* 2006a); North Carolina (Brandt 1936; HWML); Ohio (Odlaug 1936; Guberlet 1920); Oklahoma (Guberlet 1919; Trowbridge & Hefley 1934); Tennessee (Parker 1941); Texas (Harwood 1932; Hollis 1972; Mayberry *et al.* 2000; Yoder & Gomez 2007; HWML); Virginia (Campbell 1968); Wisconsin (HWML); undetermined locality (Odlaug 1936; Lemke *et al.* 2008; USNPC).

Specimens in collections: CMNPA: 1900-1617a. HWML: 20127-20131, 20957, 22683, 23424, 23425, 33140, 33141, 48349, 49007. USNPC: 50312, 50313, 51638, 51642, 51643, 82013, 84815.

Remarks: This species has been registered exclusively in North America; the only record out of this region (Kirghizia, Asia by Skarvilovich, 1950 in Yamaguti 1971) needs to be confirmed. Records of this

species from Oklahoma (representing a new species, *Gorgodera circava*) were transferred to *G. amplicava* by Harwood (1932); unfortunately, no type specimens were deposited for specific status confirmation. In the same way, specimens from Ontario, Canada (Cort 1912), Illinois (USNPC 51642, 51643 and HWML 20957), Mississippi (Brooks 1979) and Texas (Hollis 1972) were identified as *Gorgodera minima*, but Yamaguti (1971) considered this species synonym of *G. amplicava*.

Gorgodera cygnoides (Zeder, 1800)

Site of infection: urinary bladder.

Recruitment: ingestion.

Distribution: USA: New York Aquarium (USNPC).

Specimens in collections: USNPC: 36351.

Remarks: Material recorded as *Distomum cygnoides*, which is currently included in the genus *Gorgodera*. Distribution of this species is exclusively Palearctic and Oriental (Walton 1949; Prudhoe & Bray 1982); for this reason, identity of these specimens needs to be confirmed.

Gorgoderidae gen. sp.

Site of infection: urinary bladder.

Recruitment: ingestion.

Distribution: USA: California and Nebraska (HWML).

Specimens in collections: HWML: 31427, 45868, 45873.

Remarks: These specimens were found in adult stage; nevertheless, its identity remains unknown.

Gorgoderina sp.

Site of infection: kidney. Recruitment: ingestion. Distribution: USA: Nebraska (HWML). Specimens in collections: HWML: 48997.

Gorgoderina attenuata (Stafford, 1902)

Site of infection: kidney, urinary bladder.

Recruitment: ingestion.

Prevalence, mean intensity: 7.4%, 1.

Specimens deposited: CNHE: 6943.

Distribution: Canada: New Brunswick (McAlpine 1997; McAlpine & Burt 1998a; NBM); Ontario (Stafford 1903; Cort 1912; Pande 1937; USNPC); Quebec (Fantham & Porter 1948). USA: Arizona (Goldberg *et al.* 1998); Arkansas (Rosen & Manis 1976); Illinois (Andrews *et al.* 1992; USNPC); Indiana (Lank 1971); Kansas (Jinks & Johnston 1971); Massachusetts (Odlaug 1937; Rankin 1945; Goodchild 1950); Michigan (Muzzall 1991); Mississippi (Brooks 1979; HWML); Nebraska (Brooks 1976; Brooks *et al.* 2006a; HWML); North Carolina (Brandt 1936); South Carolina (USNPC); Texas (Hollis 1972; Mayberry *et al.* 2000); Virginia (Britt 1947); Washington (USNPC).

Specimens in collections: HWML: 20121, 20122, 20124, 20126, 20955, 48985, 48989, 48992-94, 48996. NBM: 791, 1448, 1484. USNPC: 44421, 50314, 82014, 84803.

Remarks: Distribution of this species extends from Canada to Guatemala, parasitizing a large variety of amphibian species, but it has not been recorded in *L. catesbeianus* from Mexico and Guatemala (Muzzall *et al.* 2001; Bolek & Coggins 2003; Mata-López *et al.* 2005; Pérez-Ponce de León *et al.* 2007).

Gorgoderina bilobata Rankin, 1937

Site of infection: urinary bladder.

Recruitment: ingestion.

Distribution: USA: Georgia (Parker 1941); Virginia (Campbell 1968).

No specimens in collections.

Remarks: This species has also been found in Caudata (*Pseudotriton* spp. and *Notophthalmus*) and other species of anurans (Yamaguti 1971; Mata-López *et al.* 2005).

Gorgoderina simplex (Looss, 1899)

Site of infection: urinary bladder.

Recruitment: ingestion.

Distribution: Canada: New Brunswick (McAlpine 1997; McAlpine & Burt 1998a; NBM); Ontario (Stafford 1903; Cort 1912; CMNPA; USNPC). USA: Michigan (Najarian 1955); Nebraska (Brooks 1976; Brooks *et al.* 2006a); undetermined locality (Bensley 1897).

Specimens in collections: CMNPA: 1900-1607, 1900-1617b. HWML: 20117, 20120. NBM: 784, 785, 788, 789, 790, 1395, 1396, 1486. USNPC 50315, 51645.

Remarks: This species was originally named *Gorgodera simplex* (Looss 1899a, 1899b) based on Canadian specimens and transferred to *Gorgoderina* by Looss (1905) who established it as the type species of the genus. Material identified by Bensley (1897) as *Distoma cygnoides* var. B was proposed as synonym of *G*. *simplex* by Looss (1905). Pande (1937) and Kaw (1950) considered this species as member of the genus *Phyllodistomum*.

Haematoloechus sp.

Site of infection: lungs.

Recruitment: ingestion.

Distribution: USA: California (HWML); Louisiana (USNPC); Nebraska (HWML); North Carolina (Brandt 1936); Texas (Knight *et al.* 1965; Morrison 1966); West Virginia (NBM).

Specimens in collections: HWML: 19716, 19840, 31435, 45945. NBM: 1372. USNPC: 59141, 59175, 59180.

Remarks: To date, 8 species of *Haematoloechus* have been recorded as parasites of *L. catesbeianus*, all of them recorded within its native distribution range.

Haematoloechus breviplexus Stafford, 1902

Site of infection: lungs.

Recruitment: ingestion.

Distribution: Canada: Ontario (Stafford 1905; Cort 1915; CMNPA); Quebec (Fantham & Porter 1948). USA: Arizona (Snyder & Tkach 2001); Arkansas (Rosen & Manis 1976; USNPC); Illinois (USNPC); Indiana (Lank 1971; Whitehouse 2002); Kentucky (Whitehouse 2002); Louisiana (Bennett 1938; USNPC; HWML); Mississippi (Clark & Longest 1969; Brooks 1979; HWML); Nebraska (Brooks *et al.* 2006a; HWML); Nevada (Babero & Golling 1974); North Carolina (Brandt 1936); New Mexico (Dronen 1977); Oklahoma (Cort 1915;

Trowbridge & Hefley 1934); South Carolina (USNPC); Texas (Knight *et al.* 1965; Slagle 1966; Hollis 1972; Underwood & Dronen 1977; Mayberry *et al.* 2000; Yoder & Gomez 2007; HWML); Virginia (Campbell 1968).

Specimens in collections: CMNPA: 1900-1621. HWML: 20293, 21957, 22681, 48350. USNPC: 50473, 82015, 84816, 95563.

Remarks: Some specimens identified by Cort (1915) as *H. breviplexus* were re-determined by León-Règagnon *et al.* (2005) as *H. floedae*. Material from Quebec was included into the genus *Pneumobites*, a synonym of *Haematoloechus* (Yamaguti 1971).

Haematoloechus buttensis Ingles, 1936

Site of infection: lungs. Recruitment: ingestion. Distribution: USA: Nevada (Babero & Golling 1974). No specimens in collections.

Haematoloechus coloradensis (Cort, 1915)

Site of infection: lungs.

Recruitment: ingestion.

Prevalence, mean intensity, and range: 3.7%, 0.11, 3.

Specimens deposited: CNHE: 6944.

Distribution: USA: Nebraska (Bolek & Janovy 2007).

Specimens in collections: HWML: 48480.

Remarks: *Haematoloechus coloradensis* is found from the western United States to Central Mexico and the Pacific coast as the result of expansion and contraction of distribution ranges of Nearctic fauna following geologic events from the early Eocene to the Pleistocene (León-Règagnon & Brooks 2003).

Haematoloechus complexus (Seely, 1906)

Site of infection: lungs.

Recruitment: ingestion.

Distribution: USA: Nebraska (Bolek & Janovy 2007; HWML); South Carolina (USNPC).

Specimens in collections: HWML: 15298, 15299, 48481. USNPC: 84805.

Remarks: *Haematoloechus complexus* has shown some specificity to leopard frogs (León-Règagnon & Brooks 2003); records in American bullfrogs may be considered as accidental infections.

Haematoloechus floedae Harwood, 1932

Site of infection: lungs.

Recruitment: ingestion.

Distribution: USA: Arizona (Mayberry *et al.* 2000; Snyder & Tkach 2001); California (León-Règagnon *et al.* 2005); Florida (Manter 1938); Georgia (León-Règagnon & Brooks 2003); Nebraska (HWML); Texas (Harwood 1932; Jacobs & Morrison 1966);

Specimens in collections: CNHE: 4663, 4664; HWML: 1251, 21945. USNPC: 30879 (holotype), 84804, 91507.

Remarks: Some authors consider *H. floedae* as junior synonym of *H. breviplexus* (Odening 1960; Kennedy 1981); however, according to morphological and molecular data, *H. floedae* is a valid species (León-Règagnon & Brooks 2003). Voucher specimens from *L. catesbeianus* identified as *H. breviplexus* by Snyder and Tkach (2001), as well as specimens deposited at the USNPC (84804), were re-assigned to *H. floedae* (León-Règagnon & Brooks 2003).

Haematoloechus lobatus (Seno, 1907)

Site of infection: lungs.

Recruitment: ingestion.

Distribution: Japan: Chiba and Kagagua Prefectures (Uchida & Itagaki 1976). Korea: Chonnam-Kohung Podu (Kim *et al.* 1992).

No specimens in collections.

Remarks: This species was described originally as *Pneumonoeces lobatus*, but the original description lacks morphological characters (Seno 1907). Uchida and Itagaki (1976) redescribed in detail this species as parasite of *Lithobates catesbeianus*.

Haematoloechus longiplexus Stafford, 1902

Site of infection: lungs.

Recruitment: ingestion.

Distribution: Canada: New Brunswick (McAlpine 1997a, 1997b; 1998; McAlpine & Burt 1998a; NBM); Nova Scotia (Stafford 1903); Ontario (Stafford 1903; Cort 1915; CMNPA); Quebec (Stafford 1903). Cuba (Odening 1968). USA: Arizona (Goldberg *et al.* 1998; Snyder & Tkach 2001; USNPC); Arkansas (Rosen & Manis 1976); California (Shields 1987; Goldberg & Bursey 2002a); Connecticut (Brooks, 1976); Georgia (HWML); Idaho (Waitz 1961; Waitz 1962); Illinois (Cort 1915); Indiana (Cort 1915; Lank 1971); Iowa (Ulmer 1970; Cain & French 1975); Kentucky (Whitehouse 2002); Louisiana (Bennett 1938); Michigan (Krull 1932; Muzzall 1991; Najarian 1955); Mississippi (Clark & Longest 1969); Nebraska (Brooks 1974; Brooks 1976; Snyder & Tkach 2001; León-Règagnon & Brooks 2003; Brooks *et al.* 2006a; Bolek & Janovy 2007; HWML; USNPC); Nevada (Babero & Golling 1974); North Carolina (Brandt 1936); Ohio (Ashton & Rabalais 1978; Bursey & DeWolf 1998); Oklahoma (Trowbridge & Hefley 1934; Brooks 1979); Oregon (HWML); Texas (Harwood 1932); Washington (Schell 1965); West Virginia (USNPC).

Specimens in collections: CMNPA: 1900-1624, 1900-1625, 1900-1626, 1900-1631, 1900-1634. HWML: 15282, 15301, 15304, 19843, 20144-20150, 20939, 21947, 23255, 48432, 48482. NBM: 793, 1244, 1245, 1248, 1249, 1251, 1460, 1479, 3523-3526. USNPC: 79466, 87069, 91244, 91509, 91510, 102161, 102175.

Remarks: León-Règagnon *et al.* (1999) synonymyzed *H. macrorchis* with *H. longiplexus*, but molecular evidence provided by Snyder & Tkach (2001) and León-Règagnon and Brooks (2003) support that they are independent species with very similar morphology.

Haematoloechus medioplexus Stafford, 1902

Site of infection: lungs.

Recruitment: ingestion. Distribution: Canada: Quebec (Fantham & Porter 1948). USA: Iowa (Cain & French 1975). No specimens in collections.

Remarks: Some species parasitizing *Lithobates* spp. in Mexico were recorded as *H. medioplexus*; further, they were re-determined as *H. danbrooksi* (León-Règagnon & Paredes-Calderón 2002). This species is a second intermediate host specialist, parasitizing only anisopteran odonates (Krull 1930; Snyder & Janovy 1994).

Haematoloechus parviplexus (Irwin 1929)

Site of infection: lungs.

Prevalence, mean intensity, and intensity range: 33.3%, 9, 1-28.

Specimens deposited: CNHE: 6945.

Recruitment: ingestion.

Distribution: USA: Connecticut (Brooks 1976); Louisiana (Bennett 1938; USNPC); Michigan (Muzzall 1991); Nebraska (Brooks 1974; Brooks 1976; León-Règagnon & Brooks 2003; León-Règagnon *et al.* 2005; Brooks *et al.* 2006a; Bolek & Janovy 2007); Nevada (Babero & Golling 1974); Washington (USNPC).

Specimens in collections: CNHE: 4405. HWML: 20142, 20143, 20753, 21660, 48433, 48483. USNPC: 75445, 81467, 95564, 102174.

Remarks: Kennedy (1981) proposed the synonymy of *H. parviplexus* with *H. varioplexus*; however, León-Règagnon and Brooks (2003) and León-Règagnon *et al.* (2005) supported the validity of this species based on molecular and morphological evidence.

Haematoloechus variegatus (Rudolphi, 1819)

Site of infection: lungs.

Recruitment: ingestion.

Distribution: Canada: Ontario (Stafford 1900). USA: New York Aquarium (USNPC).

Specimens in collections: USNPC: 35195.

Remarks: Material deposited in USNPC as *Distomum variegatum* Rudolphi; later this species was transferred to the genus *Haematoloechus* (Looss). This is a typical European species (Yamaguti 1971). Identification of this material needs to be revised.

Haematoloechus varioplexus Stafford, 1902

Site of infection: lungs.

Recruitment: ingestion.

Distribution: Canada: New Brunswick (McAlpine 1997a, 1997b; 1998; McAlpine & Burt 1998a; NBM); Ontario (Stafford 1900); Quebec (Stafford 1903). USA: Louisiana (USNPC); Nebraska (Snyder & Tkach 2001; Bolek & Janovy 2007); Virginia (Campbell 1968).

Specimens in collections: NBM: 792, 797, 798, 1257-1259, 1496-1500, 1503-1505, 3536-3538. USNPC: 84817, 91515.

Remarks: Many species of *Haematoloechus* were synonymyzed with *H. varioplexus* by Kennedy (1981). Based on morphological and molecular evidence, León-Règagnon *et al.* (2005) re-established the validity of these species and the close phylogenetic relationships of *H. varioplexus* with *H. parviplexus* and *H. colorandensis*.

Haematoloechus viguerasi Martinez, Coy-Otero & Ventosa, 1982

Site of infection: lungs.

Recruitment: ingestion.

Distribution: Cuba: Havana (Martínez et al. 1982; IES).

Specimens in collections: IES: 12.4075 (holotype), 12.4076 (paratypes).

Remarks: This species closely resembles to *H. breviplexus* and *H. floedae*; however, it can be distinguished from *H. breviplexus* by the position of the acetabulum and uterus, as well as form and size of testes; from *H. floedae* is differentiated by body lenght/width and oral sucker/acetabulum ratios, absence of esophagus, ovary shape, and distribution of vitelline glands (Martínez *et al.* 1982).

Halipegus sp.

Site of infection: Eustachian tube, stomach.

Recruitment: ingestion.

Distribution: USA: Arkansas (Rosen & Manis 1976); Louisiana (USNPC); Michigan (Muzzall 1991); Nebraska (HWML).

Specimens in collections: HWML: 19715. USNPC: 84818.

Remarks: Zelmer and Brooks (2000) restricted the genus *Halipegus* to those species lacking genital sacs, permanent sinus organs, and well-developed hermaphroditic ducts.

Halipegus amherstensis Rankin, 1944

Site of infection: Eustachian tube, mouth.

Recruitment: ingestion.

Distribution: USA: Massachusetts (Rankin 1944b; Rankin 1945).

Specimens in collections: USNPC: 36882 (holotype), 36883 (paratype).

Remarks: McAlpine and Burt (1998b) reassigned two specimens from the type material of *H*. *amhertensis* to *H. occidualis*.

Halipegus eccentricus Thomas, 1944

Site of infection: Eustachian tubes.

Recruitment: ingestion.

Distribution: USA: Michigan (Thomas 1939; Muzzall 2005, in tadpoles); Nebraska (Brooks *et al.* 2006a).

Specimens in collections: USNPC: 9203 (holotype and paratypes).

Remarks: This species was described by Thomas (1939) based on immature specimens parasitizing L. *catesbeianus*, but the material was deposited in USNPC as parasite of L. *clamitans*. McAlpine and Burt (1998b) considered this species a junior synonym of *H occidualis*, but Zelmer and Esch (1999) and McAlpine (2006) rejected this suggestion.

Halipegus occidualis Stafford, 1905

Site of infection: Eustachian tubes, mouth, stomach.

Recruitment: ingestion.

Distribution: Canada: New Brunswick (McAlpine 1997a, 1997b; 1998; McAlpine & Burt 1998b; NBM); Ontario (Stafford 1905; CMNPA). USA: Massachusetts (McAlpine & Burt 1998b); Nebraska (Brooks 1976); Pennsylvania (USNPC).

Specimens in collections: CMNPA: 1900-1629 (type). HWML: 20098-20101. NBM: 759-761, 1242, 1243, 1254, 1255. USNPC: 31124, 36883.02.

Remarks: This digenean species has been recorded from southern Canada, across USA to Mexico, in different species of amphibians (Zelmer & Brooks 2000).

Halipegus ovocaudatus (Vulpian, 1860)

Site of infection: Eustachian tube, tongue.
Recruitment: ingestion.
Distribution: Canada: Ontario (Stafford 1900).
No specimens in collections.
Remarks: According with Stunkard (1973), *H. ovocaudatus* is distributed exclusively in Europe.

Langeronia macrocirra Caballero & Bravo, 1949

Site of infection: intestine.

Recruitment: ingestion. **Distribution:** USA: Nevada (Babero & Golling 1974). **No specimens in collections.**

Remarks: These specimens were originally recorded as *Loxogenes provitellaria*, which is considered synonym of *L. macrocirra* by Ubelaker (1965). Recently, Martínez- Salazar (2004) re-examined type material of *L. provitellaria*, confirming Ubelaker's proposal.

Levinseniella ophidea Nicol, Demaree & Wootton, 1985

Site of infection: intestine.

Recruitment: ingestion.

Distribution: USA: California (Nicol et al. 1985; Goldberg & Bursey 2002a).

No specimens in collections.

Remarks: Nicol *et al.* (1985) distinguished *L. ophidea* from other members of the genus because it uses *L. catesbeianus* as the definitive host. This same authors recorded leeches as a second intermediate host for this species.

Loxogenes sp.

Site of infection: pylorus. Recruitment: ingestion. Distribution: Canada: Ontario (CMNPA). Specimens in collections: CMNPA: 1900-1619.

Loxogenes arcanum (Nickerson, 1900)

Site of infection: liver (immature cyst), duodenum, pyloric caeca, intestine,

Recruitment: ingestion.

Distribution: Canada: New Brunswick (McAlpine 1997); Ontario (Stafford 1900; Stafford 1905; CMNPA). USA: Louisiana (USNPC; HWML); Michigan (Muzzall 1991); South Carolina (USNPC).

Specimens in collections: CMNPA: 1900-1711. HWML: 22234-22242, 22354, 22684, 22697. USNPC: 84807, 84819.

Remarks: *Loxogenes arcanum* was originally described as *Distomum arcanum* by Nickerson (1900) from an undetermined North American frog species (distinct of *L. catesbeianus*) and later transferred into the genus *Loxogenes* (Stafford 1905). It has been recorded from other North American (McAlpine & Burt 1998a) and tropical frog species (Goldberg & Bursey 2007).

Loxogenoides bicolor (Krull, 1933)

Site of infection: hepatic-duodenal junction.

Recruitment: ingestion.

Distribution: USA: Carolina (USNPC); Georgia (HWML); Nebraska (Brooks *et al.* 2006a); North Carolina (Brandt 1936).

Specimens in collections: HWML: 22350, 22351. USNPC: 84808.

Remarks: This species was described as *Loxogenes bicolor* by Krull (1933), type species of the new genus *Loxogenoides* proposed by Kaw (1945).

Loxogenoides loborchis Christensen, 1981

Site of infection: liver, bile ducts.

Recruitment: ingestion.

Distribution: USA: Georgia, Kentucky, North Carolina (Christensen 1981).

Specimens in collections: HWML: 21193 (paratype). USNPC: 76097 (holotype), 76320 (paratype).

Remarks: This species and *L. bicolor* reach apparently low prevalence and intensity of infection in *L. catesbeianus*. Christensen (1981) mentioned that studies on life histories and host specifity are needed to further clarify relationships between these species.

Megalodiscus sp.

Site of infection: colon, excretory system, rectum.

Recruitment: ingestion.

Distribution: USA: Florida (Loftin 1960); Georgia (HWML); Michigan (Smith 1953).

Specimens in collections: HWML: 22244.

Remarks: This genus, erected by Chandler (1923), was considered synonym of *Diplodiscus* by Cort (1926) and Führmann (1928); however, Yamaguti (1971) enlisted *Megalodiscus* as valid genus. Specimens from Michigan were obtained experimentally and named *Megalodiscus ferrisianus* Smith, 1953, currently *nomen nudum* (Yamaguti 1971).

Megalodiscus americanus Chandler, 1923

Site of infection: rectum.

Recruitment: ingestion. Distribution: Canada: New Brunswick (McAlpine 1997). Specimens in collections: NBM.

Megalodiscus intermedius (Hunter, 1930)

Site of infection: rectum.

Recruitment: ingestion.

Distribution: USA: Louisiana (Hunter 1930; Brooks *et al.* 2006a); North Carolina (Brandt 1936). **Specimens in collections:** USNPC: 8116 (type), 45881 (paratypes), and 8117.

Remarks: Originally described as *Diplodiscus intermedius* and transferred to *Megalodiscus* by Harwood (1932). Zamparo and Brooks (2005) considered that this species could be synonym of *M. temperatus*.

Megalodiscus microphagus Ingles, 1936

Site of infection: intestine.

Recruitment: ingestion. **Distribution:** USA: Arkansas (Rosen & Manis 1976). **No specimens in collections.**

Remarks: This species was described as parasite of *Anaxyrus boreas* (Baird & Girard); besides of *L. catesbeianus*, it has been recorded from *Dicamptodon ensatus* (Eschscholtz), *Pseudacris regilla* (Baird & Girard), *Rana aurora* Baird and Girard, *Taricha granulosa* (Skilton), and *Rana cascadae* Slater (Zamparo & Brooks 2005).

Megalodiscus temperatus (Stafford, 1905)

Site of infection: intestine.

Recruitment: ingestion.

Distribution: Canada: New Brunswick (McAlpine 1997a, 1997b; McAlpine & Burt 1998a); Ontario (Stafford 1905). USA: Arkansas (Rosen & Manis 1976; USNPC); California (Ingles 1936; Goldberg & Bursey 2002a); Georgia (Parker 1941); Illinois (Andrews *et al.* 1992); Iowa (Ulmer 1970); Louisiana (USNPC); Michigan (Krull & Price 1932; Muzzall 1991; Najarian 1955); North Carolina (Brandt 1936; Brooks *et al.* 2006a; HWML); Nebraska (Brooks 1977; Brooks *et al.* 2006a); South Carolina (USNPC); Tennessee (Parker 1941); Texas (Harwood 1932; Slagle 1966; Hollis 1972; Mayberry *et al.* 2000; Yoder & Gomez 2007; HWML; USNPC); Virginia (Campbell 1968).

Specimens in collections: HWML: 4809, 4903, 20081, 20082, 20084, 33178, 48348. NBM: 1374, 1413. USNPC: 31683, 31684, 31697, 31698, 51953.02, 84284, 84285, 84809, 84820, 91245.

Remarks: Specimens of HWML (33178) were deposited as *Diplodiscus temperatus*, species currently included in *Megalodiscus*. Material from USNPC (4903) pertaining to Leidy Collection and identified as *Diplodiscus subclavatus* (Pallas) Diesing, was considered by Yamaguti (1971) synonym of *M. temperatus*.

Mesocoelium brevicaecum Ochi, 1930

Site of infection: intestine.

Recruitment: ingestion.

Distribution: Japan (Ochi 1930).

No specimens in collections.

Remarks: *Lithobates catesbeianus* has been introduced to Japan in several occasions from 1918 (Goldberg & Bursey 2002b).

Neogogatea kentuckiensis (Cable, 1935)*

Site of infection: muscles.

Recruitment: penetration **Distribution:** USA: Ohio (Myer 1960). **No specimens in collections.**

Remarks: This species was described as *Cercaria kentuckiensis* based on metacercariae obtained experimentally from tadpoles of *L. clamitans*, *L. catesbeianus* and *L. pipiens* (Cable 1935). Later, it was transferred to *Mesostephanus* by Myer (1960), and finally included into the genus *Neogogatea* (Hoffman & Dunbar 1963).

Paramphistomidae gen. sp.

Site of infection: rectum.

Recruitment: ingestion. Distribution: Canada: New Brunswick (NBM). Specimens in collections: NBM: 1419, 1420.

Pharyngostomum cordatum (Diesing, 1850)*

Site of infection: mesentery and connective tissues.

Recruitment: penetration. Distribution: Japan: Aichi Prefecture (Uchida & Itagaki 1980); Kagawa Prefecture (Uchida *et al.* 1977). No specimens in collections.

Phyllodistomum sp.

Site of infection: "cloacal bladder". Recruitment: ingestion. Distribution: Canada: Quebec (Fantham & Porter 1948). No specimens in collections.

Plagiorchis sp.

Site of infection: intestine. Recruitment: ingestion. Distribution: unknown. Specimens in collections: HWML: 42995.

Pleurogenoides sp.

Site of infection: not specified.
Recruitment: ingestion.
Distribution: USA: California, Oregon and Washington (Lehmann 1965).
No specimens in collections.

Remarks: Species of *Pleurogenoides* are parasites of reptiles (Travassos 1921); however, they can mature in *Rana* sp. (Macy 1964).

Pleurogenoides stromi Travassos, 1930

Site of infection: intestine.

Recruitment: ingestion. **Distribution:** USA: Louisiana (HWML).

Specimens in collections: HWML: 23752.

Remarks: This material needs revision; the species is typical from Europe and Asia (Rao 1977).

Proterometra albacauda Anderson & Anderson, 1967

Site of infection: esophagus, stomach.

Recruitment: ingestion.

Distribution: not specified (experimental infection by Krissinger and Mehra 1968).

No specimens in collections.

Remarks: The life cycle of this species was studied by Anderson and Anderson (1967) and Krissinger and Mehra (1968); later authors used *L. catesbeianus* as second intermediate host. This genus is a characteristic parasite of birds.

Pseudosonsinotrema catesbeianae Christian, 1971

Site of infection: duodenum.

Recruitment: ingestion.
Distribution: USA: Louisiana (USNPC); Tennessee (Christian 1971).
Specimens in collections: USNPC: 70789 (holotype), 70790 (paratype), 84821.
Remarks: This genus is parasite of reptiles and occasionally of amphibians (Yamaguti 1971).

Rauschiella linguatula (Rudolphi, 1819)

Site of infection: intestine.

Recruitment: ingestion.

Distribution: USA: Nevada (Babero & Golling 1974).

No specimens in collections.

Remarks: originally recorded as *Glypthelmins linguatula* (Rudolphi 1819), but based on morphological evidence, this species was transferred to *Rauschiella* (Razo-Mendivil *et al.* 2006).

Rauschiella proxima (Freitas, 1941)

Site of infection: no specified.

Recruitment: ingestion.

Distribution: USA: Nevada (Babero & Golling 1974).

No specimens in collections.

Remarks: originally recorded as *Glypthelmins proxima* Freitas, 1941, but this species was transferred to *Rauschiella* (Razo-Mendivil *et al.* 2006).

Ribeiroia sp.*

Site of infection: not specified.

Recruitment: ingestion.

Distribution: USA: California (Johnson *et al.* 1999; Goldberg & Bursey 2002a); western United States (Johnson *et al.* 2002).

No specimens in collections.

Remarks: Johnson *et al.* (1999; 2002) associated the presence of larvae of this parasite with malformations in host amphibians.

Strigea elegans Chandler & Rausch, 1947*

Site of infection: muscles of tadpoles.

Recruitment: penetration. **Distribution:** USA (Miller *et al.*, 1965a, 1965b, 1965c). **No specimens in collections.**

Remarks: This species is parasite of strigiformes. *Lithobates catesbeianus* and other anurans were infected with the mesocercariae of this species under experimental conditions (Miller *et al.* 1965a, 1965b, 1965c), but it has never been recorded in amphibians in the wild.

Teloporia aspidonectes (MacCallum, 1917)

Site of infection: lungs.

Recruitment: ingestion.

Distribution: USA: Michigan (Esch & Kocan 1966).

Specimens in collections: USNPC: 61205.

Remarks: This species was described as *Paramphistomum aspidonectes* by MacCallum (1921), transferred to *Opisthoporus* by Fukui (1929) and finally included in *Teloporia* by Fukui (1933). Species of the genus *Telporia* are common parasites of turtles.

Monogenea Bychowsky, 1937

Gyrodactylus sp.

Site of infection: skin.

Recruitment: contact.

Distribution: Canada: Ontario (Crawshaw 1997). USA: New Jersey (Stunkard & Dunihue 1933a, 1933b).

No specimens in collections.

Remarks: Stunkard and Dunihue (1933a, 1933b) considered that these specimens represent a new species of *Gyrodactylus*. This has also been suggested by Wootton *et al.* (1993), who pointed out that this material could pertain to *Gyrodactylus catesbeianae*.

Gyrodactylus arcuatus Bychowsky, 1933

Site of infection: skin of tadpoles.

Recruitment: contact.

Distribution: Unknown (Paetow et al. 2009).

No specimens in collections.

Remarks: In accordance with Paetow *et al.* (2009), tadpoles of this anuran species acts as accidental hosts.

Gyrodactylus catesbeianae Wootton, Ryan, Demaree, & Critchfield, 1993

Site of infection: skin of tadpoles. Recruitment: contact.

Distribution: USA: California (Wootton et al. 1993).

Specimens in collections: HWML: 39479. USNPC: 75997 (holotype), 75998 (paratype).

Remarks: In accordance with Wooten *et al.* (1993), large infestations with this gyrodactylid species resulted in abnormal growth, deformation of the tail and eventual death of the tadpoles.

Gyrodactylus jennyae Paetow, Cone, Huyse Mcaughlin & Marcogliese, 2009

Site of infection: skin of tadpoles.

Recruitment: contact.

Distribution: Unknown, but believed to be an American bullfrog farm in Missouri, USA (Paetow *et al.* 2009).

Specimens in collections: HWML: 49087.

Remarks: This species represents the fifth described from frogs and salamanders in North America, and the second for *L. catesbeianus* (Paetow *et al.* 2009).

Cestoidea Rudolphi, 1808

Cylindrotaenia americana Jewell, 1916

Site of infection: intestine.

Recruitment: ingestion.

Distribution: Canada: New Brunswick (McAlpine 1997). USA: Massachussets (Rankin 1945; Cabrera-Guzmán *et al.* 2007); Virginia (Campbell 1968; Cabrera-Guzmán *et al.*, 2007).

Specimens in collections: NBM: 1388.

Remarks: *Cylindrotaenia americana* is a western hemisphere species that infect species of several families of Anura (Goldberg & Bursey 2008).

Bothriocephalus sp.

Site of infection: intestine.

Recruitment: ingestion.

Distribution: Canada: New Brunswick (McAlpine 1997).

Specimens in collections: NBM: 1399.

Remarks: Species of this genus are typical parasites of marine and freshwater teleosts (Bray *et al.* 1994); however, some species, e.g., *Bothriocephalus acheilognathi* Yamaguti, have been recorded parasitizing amphibians and reptiles (Paredes-León *et al.* 2008).

Ophiotaenia sp.

Site of infection: intestine.

Recruitment: ingestion.

Distribution: Cuba: Provincia Habana, Provincia Pinar del Río (Martínez et al. 1982).

No specimens in collections.

Remarks: Specimens collected in Cuba were immature; this condition precluded its specific determination. To date, the only species of *Ophiotaenia* recorded in Cuba as parasite of amphibians is *Ophiotaenia bufonis* Pérez-Vigueras, 1942, in the "bufo" *Peltophryne peltocephala* (Pérez-Vigueras 1942; Freze & Rysavy 1976; de Chambrier *et al.* 2006).

Ophiotaenia gracilis Jones, Cheng & Gillespie, 1958

Site of infection: intestine.

Recruitment: ingestion.
Distribution: USA: Colorado (Buhler 1968, 1970; HWML); Virginia (Jones *et al.* 1958).
Specimens in collections: HWML: 33913, 37201.

Remarks: Ophiotaenia gracilis and Ophiotaenia magna share Lithobates catesbeianus as type host. Position of genital pore in mature proglottids (pre-equatorial in *O. magna* and equatorial in *O. gracilis*) differentiate them (Hannum 1925; Jones *et al.* 1958).

Ophiotaenia magna Hannum, 1925

Site of infection: intestine.

Recruitment: ingestion.

Distribution: USA: California (Goldberg & Bursey 2002a); Nebraka (Brooks 1976; Brooks *et al.* 2006a; HWML); Nevada (Babero & Golling 1974); Oklahoma (Hannum 1925; Trowbridge & Hefley 1934; Kuntz & Self 1944); Texas (Harwood 1932; Hollis 1972; Mayberry *et al.* 2000).

Specimens in collections: HWML: 20205. USNPC: 91249.

Remarks: see comments on O. gracilis.

Ophiotaenia saphena Osler, 1931

Site of infection: intestine.

Recruitment: ingestion.

Distribution: Canada: New Brunswick (McAlpine 1997a 1997b, 1998; McAlpine & Burt 1998a; NBM). USA: Nebraska (Brooks *et al.* 2006a); North Carolina (Brandt 1936).

Specimens in collections: NBM: 1402, 1404, 1406.

Remarks: This species is a common parasite of Lithobates clamitans (Muzzall 2005).

Proteocephalidae gen. sp.*

Site of infection: stomach.

Recruitment: ingestion.

Distribution: Canada: New Brunswick (McAlpine 1997a, 1997b; McAlpine & Burt 1998a; NBM). **Specimens in collections:** NBM: 1409.

Proteocephalus sp.

Site of infection: no specified.

Recruitment: ingestion.

Distribution: Canada: New Brunswick (McAlpine 1997). USA: California (Lehmann 1965); Nebraska (Brooks, 1976); North Carolina (Brandt 1936; cysts); Oregon (Lehmann 1965); Washington (Lehmann 1965).

No specimens in collections.

Remarks: *Proteocephalus* species are fundamentally parasites of Palaearctic fishes (de Chambrier *et al.* 2004).

Spirometra mansonoides (Mueller, 1935)*

Site of infection: femoral muscle.

Recruitment: ingestion.

Distribution: USA: Louisiana (Corkum 1966).

No specimens in collections.

Remarks: This cestode is parasite of mammals, mainly canids and felids, but it has also been registered as parasite of man (Tantaleán & Michaud 2005). Amphibians and reptiles act as intermediate hosts of this parasite (Mueller 1974).

Spirometra ranarum (Meggitt 1925)*

Site of infection: muscle.

Recruitment: ingestion.
Distribution: Canada: Quebec (Fantham & Porter 1948)
No specimens in collections.
Remarks: recorded as *Diphyllobothrium ranarum* Meggitt, 1925 (Fantham & Porter 1948).

Acanthocephala (Rudolphi, 1808)

Acanthocephalus sp.

Site of infection: intestine.

Recruitment: ingestion.

Distribution: USA: Massachusetts (USNPC).

Specimens in collections: USNPC: 38679.

Remarks: species of *Acanthocephalus* are common intestinal parasites of fishes, amphibians, and snakes (Bursey & Goldberg 2003). Members of this genus show low levels of specificity for the definitive host, e.g. *Acanthocephalus ranae* has been found in 8 genera of amphibians (Nagasawa & Egusa 1981).

Centrorhynchus sp.*

Site of infection: body cavity.

Recruitment: ingestion.

Distribution: USA: North Carolina (Brandt 1936); Texas (Hollis 1972; Mayberry *et al.* 2000); Virginia (Campbell 1968).

No specimens in collections.

Remarks: Species of genus *Centrorhynchus* include diurnal or nocturnal rapacious birds (Falconiforms or Strigiforms) as definitive hosts, and terrestrial isopods or insects, as intermediate hosts. Their various paratenic hosts (amphibians, reptiles, and mammals), play a fundamental role in their transmission to birds (Torres & Puga 1996).

Echinorhynchus sp.*

Site of infection: peritoneum.

Recruitment: ingestion.

Distribution: USA: New York Aquarium (USNPC).

Specimens in collections: USNPC: 36613.

Remarks: These acanthocephalans are common parasites of freshwater and marine fishes (Yamaguti 1963).

Fessisentis friedi Nickol, 1972

Site of infection: intestine.

Recruitment: ingestion.

Distribution: Canada: New Brunswick (McAlpine 1996, 1998; McAlpine & Burt 1998a; NBM).

Specimens in collections: NBM: 1412.

Remarks: this species is a common parasite of freshwater fishes (Choudhury & Dick 1998).

Neoechinorhynchus sp.*

Site of infection: between skin and muscles, lungs and peritoneum.

Recruitment: ingestion.

Distribution: Cuba: Provincia Pinar del Río (Martínez et al. 1982).

Specimens in collections: CZACC: 114077.

Remarks: In accordance with Amin (2002), only 1 species of the genus *Neoechinorhynchus* has been recorded parasitizing amphibians: *Neoechinorhynchus cyanophlytic* Kaw from India.

Neoechinorhynchus sp.

Site of infection: intestine.

Recruitment: ingestion.

Distribution: USA: Texas (Yoder & Gomez 2007; HWML).

Specimens in collections: HWML: 48355.

Remarks: This material can not be identified to specific level because only one specimen was collected; this suggests the accidental nature of this association (Yoder & Gomez 2007).

Neoechinorhynchus rutili (Müller, 1780)

Site of infection: intestine.

Recruitment: ingestion.

Distribution: Canada: New Brunswick (McAlpine 1996, 1997a, 1997b, 1998; McAlpine & Burt 1998a; NBM).

Specimens in collections: NBM: 1401.

Remarks: This acanthocephalan species is a common parasite of freshwater fishes (Merritt & Pratt 1964), but it can be accidental parasite of frogs and turtles (Van Cleave & Lynch 1950).

Nematoda Rudolphi, 1808

Abbreviata sp.*

Site of infection: intestine, stomach.

Recruitment: ingestion.

Distribution: Cuba: Provincia Pinar del Río (Martínez *et al.* 1982). USA: Virginia (Campbell 1968). No specimens in collections.

Remarks: Anderson (2000) pointed out that species of *Abbreviata* require insects as intermediate host. Larval specimens of this genus have been recorded from lizards, mainly in Australia and Borneo (Myers & Kuntz 1969; Goldberg *et al.* 2000) and from Cuba in 10 species of reptiles and one amphibian species (Barus 1972; Coy-Otero & Barus 1979).

Abbreviata ranae (Walton, 1931)*

Site of infection: intestine, stomach.

Recruitment: ingestion.

Distribution: USA: Columbia, Illinois, Indiana, Louisiana, South Carolina (Walton 1931).

Specimens in collections: Walton (1931) pointed out that type material was deposited in Ward Collection, University of Illinois, catalogue number 12211.

Remarks: Larval forms of this species [originally described as *Physaloptera ranae* by Walton (1931)] have been reported in frogs (Reiber *et al.* 1940; Campbell 1968; Ashton & Rabalais 1978); adults have not been described. *Physaloptera ranae* was transferred by Morgan (1941) to the genus *Abbreviata*, but Baker (1987) considered it a *species inquirendae*, because it is possible that more than one physalopterid species is envolved in these reports. On the other hand, Bursey *et al.* (2006a) pointed out that amphibians are probably intermediate hosts for species of *Abbreviata*.

Agamascaris odontocephala Steiner, 1924*

Site of infection: body cavity, liver, and stomach cysts.

Recruitment: unknown.

Distribution: USA: North Carolina (Brandt 1936); South Carolina (Walton 1933).

Specimens in collections: USNPC: 1569, 50576, 50577.

Remarks: *A. odontocephala* was described based on larval specimens collected from the stomach wall and liver of *Hyla carolinensis* (Steiner 1924).

Agamonema sp.*

Site of infection: entire body.

Recruitment: unknown.

Distribution: USA: North Carolina (Brandt 1936).

No specimens in collections.

Remarks: Definitive hosts for species of this genus are fishes; amphibians probably act as intermediate hosts.

Angiostrongylus cantonensis (Chen, 1933)

Site of infection: no specified.

Recruitment: ingestion.

Distribution: Japan (Uchida 1976; Otsuru 1977; Hasegawa & Asakawa 2004).

No specimens in collections.

Remarks: This species is parasite of rodents around the world, and can also be parasite of man (Slom *et al.* 2002).

Anisakinae gen. sp.*

Site of infection: bladder mesentery.

Recruitment: ingestion.

Distribution: USA: Oklahoma (USNPC).

Specimens in collections: USNPC: 44302.

Remarks: immature specimens. Members of Anisakinae are parasites mainly of marine mammals, turtles, fish-eating birds, and elasmobranches (Anderson 2000). Probably, *L. catesbeianus* represents an accidental host.

Aplectana sp.

Site of infection: cloaca, intestine.

Recruitment: ingestion.

Distribution: USA: Michigan (Muzzall 1991).

No specimens in collections.

Remarks: Species of the genus *Aplectana* are common intestinal parasites of reptiles and amphibians (Anderson 2000).

Aplectana cubana Barus, 1972

Site of infection: intestine.

Recruitment: ingestion.

Distribution: Cuba: Provincia Pinar del Río and Provincia Habana (Barus 1972; Martínez et al. 1982).

No specimens in collections.

Remarks: Species considered *incerta sedis* by Baker (1987).

Aplectana haematospicula Walton, 1940

Site of infection: intestine.
Recruitment: ingestion.
Distribution: Cuba: Provincia Santiago de Cuba (Martínez *et al.* 1982).
No specimens in collections.
Remarks: this species has been recorded in Cuba from 12 other amphibian host species (Walton 1940; Barus 1973; Coy-Otero & Ventosa 1984).

Ascarididae gen. sp.*

Site of infection: bladder.

Recruitment: ingestion. Prevalence, mean intensity, and intensity range: 18.5%, 0.70, 1-12. Specimens deposited: CNHE: 6950. Distribution: USA: Arizona (Goldberg *et al.* 1998). No specimens in collections.

Ascaris sp.*

Site of infection: peritoneum. Recruitment: ingestion.

Distribution: USA: Massachussetts (USNPC); New York Aquarium (USNPC).

Specimens in collections: USNPC: 34634, 34635.

Remarks: this record is doubtful. Species of *Ascaris* are exclusively parasites of mammals. Sometimes this generic name is used in place of the family or superfamily name and is sometimes used as a common term for an elongate parasitic worm.

Brevimulticaecum sp.*

Site of infection: muscle.

Recruitment: ingestion. **Distribution:** USA: Florida (Walton 1937).

No specimens in collections.

Remarks: *Lithobates catesbeianus* is considered by Bursey & Goldberg (2005) as paratenic host for this nematode.

Camallanus sp.

Site of infection: intestine.

Recruitment: ingestion.

Distribution: USA: Illinois (USNPC).

Specimens deposited: USNPC: 82017.

Remarks: Species of *Camallanus* are parasites of fishes; they are recruited by ingestion of infected copepods (Cheng 1973). This record could be accidental.

Camallanus multilineatus Kung, 1948

Site of infection: stomach.

Recruitment: ingestion. **Distribution:** UK: London Zoo (Baker 1987). **No specimens in collections.**

Remarks: Baker (1987) pointed out that despite numerous parasitological surveys on *L. catesbeianus* in its native range, this nematode species has never been found there.

Contracaecum sp.*

Site of infection: coelom, muscles, stomach.

Recruitment: ingestion.

Distribution: Cuba: Provincia Pinar del Río (Martínez *et al.* 1982). USA: California (Goldberg & Bursey 2002a; USNPC); Pennsylvania (USNPC).

Specimens in collections: USNPC: 31123, 91247.

Remarks: Life cycle of *Contracaecum* spp. includes invertebrates and fishes as intermediate hosts; records in *L. catesbeianus* are probably accidental.

Cosmocercoides sp.

Site of infection: intestine.

Recruitment: penetration.

Distribution: USA: Kansas (USNPC); Michigan (Muzzall 1991).

Specimens in collections: USNPC: 91217.

Remarks: The genus *Cosmocercoides* Wilkie comprises 17 species parasites of amphibians and reptiles from different bio-geographic Realms (Vanderburgh & Anderson 1987a, 1987b); only 2 of them have been recorded in *L. catesbeianus* (see below).

Cosmocercoides dukae (Holl, 1928)

Site of infection: intestine.

Recruitment: ingestion.

Distribution: Canada: New Brunswick (McAlpine 1997). USA: California (Lehmann 1965); Illinois (USNPC); Massachusetts (Rankin 1945); Mississipi basin (Walton 1929); North Carolina (Brandt 1936); Oregon (Lehmann 1965); Texas (Harwood 1932; Slagle 1966; Hollis 1972; Mayberry *et al.* 2000); Virginia (Campbell 1968); Washington (Lehmann 1965).

Specimens in collections: USNPC: 82018.

Remarks: This species is a common parasite of mollusks; accidentally infects frogs and salamanders (Vanderburgh & Anderson 1987a, 1987c). In accordance with these authors, many records of this species in amphibian hosts may have been confused with its closely related species *C. variabilis*.

Cosmocercoides variabilis (Harwood, 1930)

Site of infection: intestine.

Recruitment: penetration.

Prevalence, mean intensity, and range: 14.8%, 2, 1-3.

Specimens deposited: CNHE: 6946.

Distribution: Canada: New Brunswick (McAlpine & Burt 1998a); Quebec (Rau *et al.* 1978). USA: California (Ingles 1936; Lehmann 1965; Goldberg & Bursey 2002a); Illinois (Andrews *et al.* 1992); Michigan (Muzzall 1991); Mississippi (Walton 1929); North Carolina (Brandt 1936); Ohio (Bursey & DeWolf 1998); Oklahoma (Trowbridge & Hefley 1934); Oregon (Lehmann 1965). Texas (Harwood 1932; Hollis 1972; Mayberry *et al.* 2000); Virginia (Campbell 1968); Washington (Lehmann 1965).

Specimens in collections: USNPC: 91250.

Remarks: This nematode species is considered as a true parasite of amphibians (Bolek 1997).

Dioctophyma renale (Goeze, 1782)*

Site of infection: body cavity.

Recruitment: ingestion.

Distribution: Canada: Ontario (Mace & Anderson 1975).

No specimens in collections.

Remarks: This parasite has been found encapsulated in fishes; larvae can be transmitted along food chain of paratenic hosts (Karmanova 1961). Possibly American bullfrogs became infected with *D. renale* by eating parasitized frogs of different species (Mace & Anderson 1975).

Dioctophymidae gen. sp.*

Site of infection: muscles.

Recruitment: ingestion.

Distribution: USA: Nevada (USNPC).

Specimens in collections: USNPC: 66950.

Remarks: Species of this family are associated with the kidneys and alimentary tract of carnivore mammals and birds, respectively (Anderson & Bain 1982).

Dujardinascaris sp.*

Site of infection: body cavity.

Recruitment: ingestion.

Distribution: USA: North Carolina (Brandt 1936).

No specimens in collections.

Remarks: Originally recorded as *Dujardinia* by Brandt (1936). Adults of *Dujardinascaris* spp. are parasites of fishes and aquatic reptiles (Sprent 1977).

Enterobius vermicularis Linnaeus, 1758

Site of nfection: intestine.

Recruitment: ingestion.

Distribution: USA: New York Aquarium (USNPC).

Specimens in collections: USNPC: 34738.

Remarks: This identification is doubtful; *E. vermicularis* is a human parasite (Lamothe-Argumedo & García-Prieto 1988). Specimens deposited at USNPC as *Oxyuris vermicularis*.

Eustrongylides sp.*

Site of infection: body cavity attached to mesenteries, under skin.

Recruitment: ingestion.

Distribution: Cuba (Coy-Otero & Martínez 1987). USA: California (Goldberg & Bursey 2002a). **Specimens in collections:** USNPC: 44423, 72571, 91252.

Remarks: Adult forms of *Eustrongylides* inhabit the pro-ventriculus of aquatic birds, whereas the infective larval stage is found in the tissues of fishes, amphibians and reptiles (Panesar & Beaver 1979).

Eustrongylides wenrichi Canavan, 1929*

Site of infection: eye, kidneys, liver, mesentery, mouth, muscle, peritoneum, skin, spleen, stomach. **Recruitment:** ingestion.

Distribution: USA: Louisiana (Modzelewski & Culley 1974); Nevada (Babero & Golling 1974); Pennsylvania (USNPC); undetermined (Walton 1935).

Specimens in collections: USNPC: 80887 (paratype).

Remarks: Panesar & Beaver (1979) described the third larval stage of this parasite.

Falcaustra sp.*

Site of infection: musculature.

Recruitment: ingestion.

Distribution: USA: Texas (Yoder & Gomez 2007).

Specimens in collections: HWML: 48354.

Falcaustra sp.

Site of infection: intestine, stomach.

Recruitment: ingestion.

Distribution: USA: Oklahoma (Kuntz & Self 1944).

Specimens in collections: USNPC: 44288.

Remarks: There are currently 69 nominal species of *Falcaustra* that occur in the digestive tract of fishes, amphibians, and reptiles (Bursey & Goldberg 2001). Specimens were deposited as *Spironoura*, junior synonym of *Falcaustra* (Freitas & Lent 1941).

Falcaustra catesbeianae Walton, 1929

Site of infection: intestine.

Recruitment: ingestion.

Distribution: Canada: Ontario (Baker 1986a; CMNPA). Japan: Hasama Oita (Hasegawa 2006). USA: Arkansas (McAllister *et al.* 2008; USNPC); California (Goldberg & Bursey 2002a); Georgia (Reiber *et al.* 1940); Illinois (Walton 1929; Walton 1933; USNPC); Louisiana (Walton 1929; USNPC); Maryland (USNPC); Michigan (Muzzall 1991); Nevada (Babero & Golling 1974); North Carolina (Brandt 1936); Ohio (Ashton & Rabalais 1978); Oklahoma (Walton 1929; Trowbridge & Hefley 1933; Kuntz & Self 1944; USNPC); South Carolina (USNPC); Tennessee (Reiber 1941); Texas (Harwood 1932; Slagle 1966; USNPC).

Specimens in collections: CMNPA: 1985-0161. NSMT-As: 3031. USNPC: 33061, 44272, 44299, 50683, 50685, 50808, 50809, 52137, 91246, 52131, 66528, 84247, 84248, 84796, 84810, 91246.

Remarks: Specimens from USNPC 44299, 50808, 50809, 52131, 66528 were recorded as *Spironoura catesbianae*. Record from Japan is considered an introduction by American bullfrogs imported from North America (Hasegawa 2006).

Falcaustra inglisi (Anderson, 1964)

Site of infection: intestine.

Recruitment: ingestion.

Distribution: Canada: New Brunswick (McAlpine 1997); Ontario (Anderson 1964; Baker 1986a; CMNPA; USNPC).

Specimens in collections: CMNPA: 1978-1196, 1985-0162. USNPC: 60164.

Remarks: Species originally described as *Oxysomatium inglisi* (Anderson 1964) and transferred to *Falcaustra* by Baker (1980).

Filaria nitida Leidy, 1856

Site of infection: intestine.

Recruitment: vector. Distribution: USA: New York Aquarium (USNPC). Specimens in collections: USNPC: 34734. Remarks: *Species inquerendae* (Baker 1987).

Filaria quadrituberculata Leydi, 1856*

Site of infection: peritoneum and mesentery.

Recruitment: vector.

Distribution: USA: Louisiana (Walton 1927); New Jersey (Benach 1972); Oklahoma (Trowbridge & Hefley 1934; Kuntz & Self 1944).

No specimens in collections.

Remarks: Species inquirendae (Baker 1987).

Foleyella sp.

Site of infection: body cavity around the liver and spleen, intestinal mesentery, mesentery about the kidneys. **Recruitment:** vector.

Distribution: USA: New Jersey (Crans 1969; USNPC); Oklahoma (USNPC).

Specimens in collections: USNPC: 44293, 71484, 71485, 71486, 71487.

Remarks: Bartlett (1986) emended the diagnosis of *Foleyella* including only 3 species and 1 subspecies: *F. candezei* (Fraipont) Seurat; *F. furcata* (Linstow) Witenberg & Gerichter; *F. brevicauda brevicauda* Chabaud & Brygoo; *F. b. magnilarvata* Bain, and *F. philistinae* Schacher & Khalil. USNPC 71484–71487 were recorded as *Foleyella seasonalis*, which is considered *nomen nudum*.

Foleyellides sp.

Site of infection: body cavity, encysted on peritoneum and mesentery.

Recruitment: vector.

Distribution: USA: New Jersey (Crans 1969; Benach 1972, experimental); Oklahoma (Trowbridge & Hefley 1934; Kuntz & Self 1944); Texas (Yoder & Gomez 2007).

Specimens in collections: HWML: 48352.

Foleyellides americana (Walton, 1929)

Site of infection: body cavity, mesentery, bladder.

Recruitment: vector.

Distribution: USA: Texas (Hollis 1972; Mayberry *et al.* 2000); North Carolina (Brandt 1936); Virginia (Campbell 1968); Ohio (Ashton & Rabalais 1978).

No specimens in collections.

Remarks: This species was described as *Foleyella americana* (Walton 1929), and transferred to *Foleyellides* by Esslinger (1986).

Foleyellides flexicauda (Schacher & Crans, 1973)

Site of infection: blood, body cavity, mesentery.

Recruitment: vector.

Distribution: USA: New Jersey (Schacher & Crans 1973; Terwedow & Craig 1977a, 1977b; Baker 1987); experimental (Benach & Crans 1973).

Specimens in collections: USNPC: 72555 (holotype), 72556 (allotype), 72557 (male paratypes), 72558 (female paratypes), 72559 (microfilariae), 72600-72603 (vouchers).

Remarks: This species was registered as *Foleyella flexicauda* (Schacher & Crans 1973), which is considered synonym of *Foleyellides flexicauda* in accordance with Bartlett (1986) and Esslinger (1986).

Foleyellides ranae (Walton, 1929)

Site of infection: encysted in body cavity and mesentery.

Recruitment: vector.

Distribution: USA: Louisiana (Walton 1929; Walton 1935; Causey 1939a, 1939b; Kotcher 1941; Baker 1987); North Carolina (Brandt 1936).

No specimens in collections.

Remarks: This species was described as member of the genus *Foleyella* (Walton, 1929); however, Esslinger (1986) re-established the genus *Foleyellides* to include those species of *Foleyella* parasites of amphibians.

Gnathostoma spinigerum Owen, 1836*

Site of infection: muscles.

Recruitment: ingestion.

Distribution: Japan (Uchida 1975; Hasegawa & Asakawa 2004).

No specimens in collections.

Remarks: Specimens of these records were collected as larvae; frogs act as paratenic hosts of this nematode.

Gyrinicola batrachiensis (Walton, 1929)

Site of infection: intestine.

Recruitment: ingestion.

Distribution: Canada: New Brunswick (Adamson 1981a); Ontario (Adamson 1981a; Pryor & Greiner 2004; CMNPA); Quebec (Adamson 1981a; Pryor & Greiner 2004). USA: Florida (Pryor & Greiner 2004); Ohio (Pryor & Greiner 2004); undetermined (Pryor & Bjorndal 2005).

Specimens in collections: CMNPA: 1980-1671a, 1980-1671b. USNPC: 92589.

Remarks: *Gyrinicola batrachiensis* can infect the gastrointestinal tracts of anuran larvae, but it is not observed in metamorphosing tadpoles or adult frogs (Adamson 1981b; Adamson 1981c; Bursey & DeWolf 1998).

Hedruris sp.

Site of infection: not specified.

Recruitment: ingestion.

Distribution: USA: California (Lehmann 1965); New Hampshire (Muzzall 1991); Oregon (Lehmann 1965); Washington (Lehmann 1965).

No specimens in collections.

Remarks: To date, 21 nominal species of *Hedruris* have been described; they occur in the digestive tract of fishes, frogs, salamanders, lizards, and turtles (Bursey & Goldberg 2007).

Hedruris pendula (Leidy, 1851)

Site of infection: intestine.

Recruitment: ingestion.

Distribution: Canada: Ontario (Baker 1986b).

Specimens in collections: CMNPA: 1986-0003.

Remarks: *H. pendula* and *H. siredonis* are the only two valid *Hedruris* species distributed in North America (Baker 1986b) parasitizing aquatic vertebrates.

Hedruris siredonis (Baird, 1858)

Site of infection: stomach.

Recruitment: ingestion.

Distribution: USA: New Hampshire (Muzzall & Baker 1987).

No specimens in collections.

Remarks: Although this species is more frequently recorded as parasite of salamanders, infections in American bullfrogs do not appear to be accidental, and they can acquire the infection by predation on aquatic infected isopods (Muzzall & Baker 1987).

Kathlaniidae gen. sp.

Site of infection: intestine, stomach. Recruitment: ingestion. Distribution: USA: Texas (USNPC). Specimens in collections: USNPC: 31180.

Onchocercidae gen. sp.*

Site of infection: blood vessels.

Recruitment: vector.Distribution: USA: North Carolina (Brandt 1936); Virginia (Campbell 1968).No specimens in collections.Remarks: these larvae were referred to the informal collective group Microfilariae.

Oswaldocruzia sp.

Site of infection: intestine.

Recruitment: penetration.

Distribution: USA: Michigan (Ridgeway 1964; Muzzall 2005); Tennessee (USNPC).

Specimens in collections: USNPC: 42061.

Remarks: the 81 species assigned to *Oswaldocruzia* are typically found in the intestine of amphibians and reptiles (Bursey *et al.* 2006b; Durette-Desset *et al.* 2006).

Oswaldocruzia leidyi Steiner, 1924

Site of infection: intestine. Recruitment: penetration. Distribution: USA: Tennessee (USNPC).

Specimens in collections: USNPC: 42061.

Remarks: Baker (1977) considered *Oswaldocruzia leydi* Travassos, 1917 as a *nomen nudum*; Ben Slimane & Durette-Desset (1997) re-established its validity, noted that the first description of this species was provided by Steiner (1924), becoming Stainer the author of the name.

Oswaldocruzia lenteixeirai Pérez Vigueras, 1938

Site of infection: intestine.

Recruitment: penetration.

Distribution: Cuba: Provincia Santiago de Cuba and Provincia Habana (Moravec & Kaiser 1995).

Specimens in collections: USNPC: 42061.

Remarks: Besides *L. catesbeianus*, this species has been recorded from Cuba parasitizing 22 species of reptiles and 9 species of amphibians (Barus & Moravec 1967; Barus 1972; Barus 1973; Coy-Otero & Barus 1979).

Oswaldocruzia pipiens Walton, 1929

Site of infection: intestine.

Recruitment: penetration.

Distribution: Canada: New Brunswick (McAlpine 1997); Ontario (Baker 1978a). USA: California (Goldberg & Bursey 2002a; USNPC); Louisiana (USNPC); North Carolina (Brandt 1936); Oklahoma (Trowbridge & Hefley 1934); Virginia (Campbell 1968).

Specimens in collections: USNPC: 91241, 84811, 91251.

Remarks: Ben Slimane and Durette-Desset (1997) considered that *O. pipiens* constitutes a species complex inhabiting North American amphibians.

Oxysomatium sp.*

Site of infection: not specified.
Recruitment: unknown.
Distribution: USA: Oklahoma (Trowbridge & Hefley 1934).
No specimens in collections.

Oxyurida gen. sp.*

Site of infection: not specified. Recruitment: ingestion. Distribution: Canada: Ontario (CMNPA). Specimens in collections: CMNPA: 1979-0492.

Parapharyngodon bassi (Walton, 1940)

Site of infection: intestine.

Recruitment: ingestion. **Distribution:** Cuba: Provincia Habana and Provincia Santiago de Cuba (Martínez *et al.* 1982). **No specimens in collections**.

Remarks: This species has been recorded in Cuba from other amphibian hosts by Walton (1940), Barus and Moravec (1967) and Barus (1973).

Physalopteridae gen. sp.*

Site of nfection: stomach.

Recruitment: ingestion. **Distribution:** USA: Oklahoma (USNPC). **Specimens in collections:** USNPC: 44286.

Physaloptera sp.*

Site of infection: body cavity, intestinal wall, mesentery, stomach.

Recruitment: ingestion.

Distribution: USA: Arizona (Goldberg *et al.* 1998); California (Goldberg & Bursey 2002a); Indiana (Walton 1935; USNPC); North Carolina (Brandt 1936); Ohio (Ashton & Rabalais 1978); Virginia (Campbell 1968).

Specimens in collections: USNPC: 50789, 87072, 91253.

Porrocaecum sp.*

Site of infection: body cavity, intestine.

Recruitment: ingestion.

Distribution: Cuba: Provincia Pinar del Río and Provincia Santigo de Cuba (Martínez *et al.* 1982; Moravec & Kaiser 1995).

No specimens in collections.

Remarks: Larvae of this genus have been reported in 24 species of reptiles and three species of amphibians from Cuba (Barus 1972; Barus 1973; Coy-Otero & Barus 1979).

Raillietnema longicaudata (Walton, 1929)

Site of infection: intestine.

Recruitment: unknown.

Distribution: USA: Indiana (Brandt 1936; Lank 1971); North Carolina (Brandt 1936).

No specimens in collections.

Remarks: specimens recorded as *Oxysomatium longicaudata*, species included in *Raillietnema* by Baker (1985).

Rhabdias americanus Baker, 1978b

Site of infection: lungs.

Recruitment: vector.

Distribution: Canada: Quebec (Fantham & Porter 1948).

No specimens in collections.

Remarks: specimens recorded by Fantham and Porter (1948) as *Rhabdias bufonis* (Schrank, 1788); Baker (1987) included this record in *R. americanus*.

Rhabdias sp.

Site of infection: body cavity, lungs.

Recruitment: vector.

Distribution: USA: California (Lehmann 1965); North Carolina (Brandt 1936); Oklahoma (USNPC); Oregon (Lehmann, 1965); Washington (Lehmann 1965).

Specimens in collections: USNPC: 44295.

Remarks: Species of *Rhabdias* Stiles and Hassall could be identified based on their relatively strict host specificity (Martínez-Salazar 2006).

Rhabdias ranae Walton, 1929

Site of infection: lungs.

Recruitment: vector.

Prevalence, mean intensity and range: 25.9%, 2.28, 1-4.

Specimens deposited: CNHE: 6947.

Distribution: USA: Arizona (Goldberg *et al.*, 1998); Louisina (USNPC); North Carolina (Brandt 1936); Ohio (Ashton & Rabalais 1978; Bursey & DeWolf 1998); Oklahoma (Morrison 1967); Texas (Harwood 1932; Hollis 1972; Mayberry *et al.* 2000); Virginia (Campbell 1968).

Specimens in collections: USNPC: 84812.

Remarks: In accordance with Kuzmin *et al.* (2003), this species is restricted to North American frogs; however, it is more common in hosts from northern United States and Canada.

Spinitectus sp.

Site of infection: stomach.

Recruitment: ingestion.

Distribution: USA: Oklahoma (Trowbridge & Hefley 1934); Tennessee (Christian 1970); South Carolina (USNPC).

Specimens in collections: USNPC: 84800.

Remarks: According to Moravec (1998), species of this genus mainly parasitize marine and freshwater fishes; only four species have been recorded in amphibian hosts (Baker 1987).

Spinitectus gracilis Ward & Magath, 1917

Site of infection: intestine, stomach.

Recruitment: ingestion.

Prevalence, mean intensity, and intensity range: 25.9%, 6.43, 2-12.

Specimens deposited: CNHE: 6948.

Distribution: USA: Michigan (Muzzall 1991); Oklahoma (Kuntz & Self 1944; USNPC).

Specimens in collections: USNPC: 44274.

Remarks: This species rarely parasitizes amphibian hosts, and is unlikely that its life cycle ends successfully in this group of vertebrates (Baker 1987).

"Spiroptera mugientis" von Linstow, 1898*

Site of infection: intestine, stomach.

Recruitment: ingestion.

Distribution: USA: undetermined (Walton, 1935).

No specimens in collections.

Remarks: Walton (1935) refers the uncertain position of this species; Yamaguti (1961) considered *Spiroptera* synomym of *Acuaria* (in part) and *Spirura* (in part).

Spiroxys sp.*

Site of infection: intestine, mesentery.

Recruitment: ingestion.

Prevalence, mean intensity, and intensity range: 3.7%, 0.07.

Specimens deposited: CNHE: 6949.

Distribution: USA: Arizona, Michigan (Muzzall 1991); North Carolina (Brandt 1936); Texas (Yoder & Gomez 2007; HWML)

Specimens deposited: HWML: 48353.

Spiroxys contorta (Rudolphi, 1819)

Site of infection: intestine.

Recruitment: ingestion.

Distribution: Canada: New Brunswick (McAlpine 1997; NBM).

Specimens in collections: NBM: 1833.

Remarks: This species is a common parasite of turtles (Yamaguti 1961). Record in *L. catesbeiana* is accidental.

Spiroxys constricta (Leidy, 1856)

Site of infection: stomach.

Recruitment: ingestion.

Distribution: USA: Louisiana (USNPC).

Specimens in collections: USNPC: 84813.

Remarks: fishes, tadpoles and frogs serve as second intermediate hosts for *S. constricta*, as well as *S. contorta* (Hedrick 1935).

Strongyluris ranae Reiber, Byrd & Parker, 1940

Site of infection: intestine.
Recruitment: ingestion.
Distribution: USA: Georgia (Reiber *et al.* 1940; Baker 1987).
Specimens in collections: USNPC: 9383 (holotype).
Remarks: This species has been exclusively found in *L. catesbeiana*.

Discussion

All helminth species found in Pawnee Lake, Nebraska, have previously been reported for this host species. However, this is the first report of the nematode *S. gracilis* for this locality; its presence in this host species is considered an accidental infection (Baker 1987), due to the overlapping ecological niche of fishes and bull frogs. Helminth richness in this host species was 9, with 5 species of nematodes and 4 of digeneans. Only 2 of the 9 species were collected as larval forms, which indicate the important role of this frog as definitive host in Pawnee Lake. Helminth taxa were collected from 5 sites within the hosts, with lungs being the most parasitized (3 taxa). The most frequent mode of parasite recruitment was ingestion of intermediate hosts; thus, 56% of the species were recruited through the food web. In contrast, 22% of helminth species were acquired by skin penetration, while 22% (*G. attenuata* and *G. quieta*) enter to the host using both vias (ingestion or penetration) (Yamaguti 1975; Anderson 2000).

Ecological parameters of infection recorded in *L. catesbeianus* from Pawnee Lake were low, similarly to those recorded for other species of amphibians in general (Muzzall 1991; McAlpine 1997; Goldberg *et al.*, 2000; Muzzall *et al.*, 2001; Goldberg *et al.*, 2001; Bolek & Coggins 2003; Goldberg *et al.*, 2002; Paredes-Calderón *et al.*, 2004; Cabrera-Guzmán 2007). Vagility and feeding habits of hosts, as well as particular ecological conditions of each locality, have been used to explain these low levels of infection.

Because it is an opportunistic and voracious predator and has high adaptability, *L. catesbeianus* is considered to be one of the most successful species in the world by the Global Invasive Species Database (2009). As so, populations of this anuran species are currently established in various parts of the world, but its helminth fauna has not been exhaustively studied. Moreover, most of the species that have been found parasitizing this amphibian represent point localityl records; only two studies about its parasites cover states or regions (Lank 1971; Babero & Golling 1974). Prior to this study, the most comprehensive compilation about its parasite fauna was made in the United States (Andrews *et al.* 1992); this compilation included 95 helminth species (one monogenean, 51 digeneans, seven cestodes, one acanthocephalan, and 35 nematode species). As a result of our survey, and by the addition of records published or included in scientific collections, the number of helminth taxa known for this host in its native and introduced range of distribution increased to 159 (67% more); these records come from 6 countries (Canada, Cuba, Japan, Korea, United Kingdom, and USA), most of them from USA (121), followed by Canada (44). Three of the 159 species registered in this amphibian host have been considered *species inquirendae*; seven more were recorded experimentally. In addition, we found 7 unidentified records in the USNPC; their inclusion in the list of helminth taxa parasitizing *L. catesbeianus* is pending of a more precise determination.

As in other amphibian hosts, species richness is influenced by local availability of helminth species and their possibility of colonization; however, the presence of most of the helminth species is due to host diet (117 species are recruited through this via while 22 enter by penetration, and 11 more via vector transmission). Infections by accidental parasites in American bullfrogs (at least 9 species), are result of the generalist predator condition of this host species. For example, accidental infection by *G. spinigerum* in Japanese bullfrogs can be explained by ingestion of natural intermediate (copepods) or paratenic hosts (fishes) (Miyazaki 1991).

The helminth fauna of this host throughout its range is composed of 2 groups of species: generalists (comprised of helminth species that commonly occur in other groups of animals but can also infect amphibians, or that use *L. catesbeianus* as a paratenic host), and specialists (comprised of species with an "historic relationship" with the amphibian hosts, sensu Brooks & McLennan 2003). In the first group are genera such as *Alaria* and *Clinostomum*, represented by 3 species each; among specialists to anurans, two other digenean genera, *Haematoloechus* and *Halipegus*, are widely represented in the helminthological record of this frog, with 13 and 5 species, respectively, although some of the species of these genera more commonly parasitize other frog groups and are rarely found in *L. catesebeianus*, for example, *H. complexus* or *H. medioplexus*, that are more frequently found in leopard frogs (León-Règagnon 2003).

Of the 159 taxa recorded infecting American bullfrogs worldwide, only 16 species have been added to their helminth fauna from localities outside of its native distribution range. Although numerous, the helminthological record of this host species will increase after further inventory work, since this species has been introduced at least to 15 countries and helmith records come from only 4 of them (Cuba, Japan, Korea, and

United Kingdom). On the other hand, only one record of colonization of helminths from *Lithobates catesbeianus* to local amphibian fauna has been documened (León-Règagnon *et al.* 2005), but considering that host switching is a relatively frequent event in parasite evolution (Brooks & Ferrao 2005; Brooks *et al.* 2006b), it is imperative to evaluate the potential impact of introducing American bullfrogs (or any species) and their parasites, not only helminths, but also other emerging pathogens (Garner *et al.* 2006) into new environments, before doing so.

To the best of our knowledge, the only other species of anuran host that has been intensively studied from a helminthological point of view is the cane toad *Rhinella marina* (L.). Even though the natural and introduced distribution of cane toad is greater than those of the American bullfrog (*R. marina* is naturally distributed in 19 American countries and has been introduced to other 38 around the world), this bufonid species is parasitized by only 113 helminth species along its natural and introduced distribution range (Espinoza-Jiménez *et al.* 2007). Differences between helminth fauna harbored by both host species could be attributed to the particular biological characteristics of each one (e.g., *L. catesbeianus* spends time both on land and in water while *R. marina* usually stay on dry land and reproduce in any shallow water), or a merely effect of a distinct sampling effort.

Acknowledgments

We thank Hideo Hasegawa, Donald McAlpine, Luisa Ventosa, Nayla García, Judith Price, Jean-Marc Gagnon, and Georgina Ortega-Leite for providing us with some bibliographic references and/or information about scientific collections. Agustín Jiménez and Matt Bolek for helping us in field and laboratory work. Elizabeth Martínez and Ulises Razo for their help with the identification of some specimens. Special thanks to Scott L. Gardner by allowing us the facilities of Harold W. Manter Laboratory of Parasitology, University of Nebraska, Lincoln to conduct the hosts revision; amphibians were collected under permission number 164 issued by Nebraska Game and Parks Commission, Scientific Collecting Master Permit to Scott L. Gardner; R.M.L. thanks the support of the Dirección General de Asuntos del Personal Académico, Universidad Nacional Autónoma de México, through a PROFIP postdoctoral scholarship.

References

- Adamson, M.L. (1981a) *Gyrinicola batrachiensis* (Walton, 1929) n.comb. (Oxyuroidea: Nematoda) from tadpoles in eastern and central Canada. *Canadian Journal of Zoology*, 59, 1344–1350.
- Adamson, M.L. (1981b) Development and transmission of *Gyrinicola batrachiensis* (Walton, 1929) Adamson, 1981 (Pharyngodonidae: Oxyuroidea). *Canadian Journal of Zoology*, 59, 1351–1367.
- Adamson, M.L. (1981c) Seasonal changes of *Gyrinicola batrachiensis* (Walton, 1929) in wild tadpoles. *Canadian Journal of Zoology*, 59, 1377–1386.
- Augustine, D.L. & Uribe, C. (1927) Alaria arisaemoides n. sp. a trematode from Vulpes fulva. Parasitology, 19, 236.
- Amin, O. (2002) Revision of *Neoechinorhynchus* Stiles & Hassall, 1905 (Acanthocephala: Neoechinorhynchidae) with keys to 88 species in two subgenera. *Systematic Parasitology*, 53, 1–18.
- Anderson, R.C. (1964) Oxysomatium inglisi n. sp. (Nematoda: Cosmocercidae) from the bullfrog (Rana catesbeiana). Canadian Journal of Zoology, 42, 255–257.
- Anderson R.C. (2000) Nematode parasites of vertebrates: Their development and transmission. CABI Publishing, Wallingford, U.K. 521 pp.
- Anderson, M.G. & Anderson, M.A. (1967) The life histories of *Proterometra albacauda* and *Proterometra septimae*, spp. n. (Trematoda: Azygiidae) and a redescription of *Proterometra catenaria* Smith, 1934. *Journal of Parasitology*, 53, 31–37.
- Anderson, R.C. & Bain, O. (1982) Keys to the genera of the Superfamilies Rhabditoidea, Dioctophymatoidea, Trichinelloidea and Muspiceoidea. *In:* Anderson, R.C., Chabaud, A.G., & Willmott, S. (Eds.) *CIH keys of the nematode parasites of vertebrates No.9*. Commonwealth Agricultural Bureaux, Farnham Royal, Bucks, England, pp. 1-26.
- Andrews, K.D., Lampley, R.L., Gillman, M.A., Corey, D.T., Ballard, S.R., Blasczyk, M.J. & Dyer, W.G. (1992) Helminths of *Rana catesbeiana* in Southern Illinois with a checklist of helminths in bullfrogs of North America. *Transactions of the Illinois State Academy of Science*, 85, 147–172.

Ashton, A.D. & Rabalais, F.C. (1978) Helminth parasites of some anurans of Northwestern Ohio. Proceedings of the Hel-

minthological Society of Washington, 45, 141–142.

- Babero, B.B. & Golling, K. (1974) Some helminth parasites of Nevada bullfrogs, *Rana catesbeiana* Shaw. *Revista de Biología Tropical*, 21, 207–220.
- Baer, J.G. (1933) Note sur un noveaux trématode, *Clinostomum lophophallum* sp. nov. avec quelques considerations génerales sur la famille des Clinostomidae. *Revue Suisse de Zoologie*, 40, 317–342.
- Baird, W. (1858) Descriptions of five new species of Entozoa. *Proceedings of the Zoological Society of London*, 26, 224–225.
- Baker, M.R. (1977) Redescription of *Oswaldocruzia pipiens* Walton, 1929 (Nematoda: Trichostrongylidae) from amphibians of eastern North America. *Canadian Journal of Zoology*, 55, 104–109.
- Baker, M.R. (1978a) Transmission of *Cosmocercoides dukae* (Nematoda: Cosmocercoidea) to amphibians. *Journal of Parasitology*, 64, 765–766.
- Baker, M.R. (1978b) Morphology and taxonomy of *Rhabdias* spp. (Nematoda: Rhabdiasidae) from reptiles and amphibians of southern Ontario. *Canadian Journal of Zoology*, 56, 2127–2141.
- Baker, M.R. (1980) Reclassification of *Oxysomatium inglisi* Anderson, 1964 and *Aplectana gigantica* Olsen, 1938 (Nematoda: Cosmocercoidea) from North American frogs. *Systematic Parasitology*, 1, 245–253.
- Baker, M.R. (1985) *Raillietnema longicaudata* (Walton, 1929) n.comb. (Nematoda: Cosmocercidae) from North American frogs. *Proceedings of the Helminthological Society of Washington*, 52, 76–79.
- Baker, M.R. (1986a) *Falcaustra* species (Nematoda: Kathlaniidae) parasitic in turtles and frogs in Ontario. *Canadian Journal of Zoology*, 64, 228–237.
- Baker, M.R. (1986b) Revision of *Hedruris* Nitzsch (Nematoda: Habronematoidea) from aquatic vertebrates of North America. *Canadian Journal of Zoology*, 64, 1567–1572.
- Baker, M.R. (1987) Synopsis of the Nematoda parasitic in amphibians and reptiles. *Memorial University of Newfoundland Occasional Papers in Biology*, 11, 1–325.
- Bartlett, C.M. (1986) The reptilian filarioid genus *Foleyella* Seurat, 1917 (Onchocercidae: Dirofilariinae) and its relationship to other dirofilariine genera. *Systematic Parasitology*, 9, 43–56.
- Barus, V. (1972) Nematodes parasitizing hosts of the genus *Eleutherodactylus* (Amphibia) from Cuba. *Vestnik Ceskoslov*enske Sponecnosti Zoologicke, 36, 161–168.
- Barus, V. (1973) Nematodes parasitizing hosts of the genus Bufo (Amphibia) in Cuba. Folia Parasitologica, 20, 29-39.
- Barus, V. & Moravec, F. (1967) Systematic studies of parasitic worms, found in the hosts *Lepisoteus tristoechus* (Ginglymodi, Lepisosteidae) and *Hyla insulsa* (Ecaudata: Hylidae) from Cuba. *Vestnik Ceskoslovenske Sponecnosti Zoologicke*, 31, 1–14.
- Beaver, P.C. (1929) Studies on the development of Allassostoma parvum Stunkard. Journal of Parasitology, 16, 13-23.
- Ben Slimane, B. & Durette-Desset, M-C. (1997) Révision du genre *Oswaldocruzia* (Nematoda, Trichostrongylina, Molineoidea) en zone néarctique avec description de cinq nouvelles espèces. *Zoosystema*, 19, 61–79.
- Benach, J.L. (1972) Studies on amphibians filariasis: relationships of *Foleyella* sp. (Filarioidea: Nematoda) from the bullfrog, *Rana catesbeiana* and *Culex territans* (Culicidae: Diptera). *Dissertation Abstracts International*, 32B, 5231.
- Benach, J.L. & Crans, W.J. (1973) Larval development and transmission of *Foleyella flexicauda* Schacher and Crans, 1973 (Nematoda: Filarioidea) in *Culex territans* (Culicidae: Diptera). *Journal of Parasitology*, 59, 797–800.
- Bennett, H.J. (1938) A partial check list of the trematodes of Louisiana vertebrates. Louisiana Academy of Science, 4, 178–181.
- Bensley, R.R. (1897) Two forms of Distomum cygnoides. Zentralblatt fur Bakteriologie Parasitenkunde und Infections Krankheitem, 21, 326–331.
- Bolek, M.G. (1997) Seasonal ocurrence of *Cosmocercoides dukae* and prey analysis in the blue-spotted salamander, *Ambystoma laterale*, in southeastern Wisconsin. *Journal of the Helminthological Society of Washington*, 64, 292–295.
- Bolek, M.G. & Coggins, J.R. (2003) Helminth community structure of sympatric eastern American toad, *Bufo americanus americanus*, northern leopard frog, *Rana pipiens*, and blue-spotted salamander, *Ambystoma laterale*, from southeast-ern Wisconsin. *Journal of Parasitology*, 89, 673–680.
- Bolek, M.G., & Janovy, J. Jr. (2007) Evolutionary avenues for and constraints on the transmission of frog lung flukes (*Haematoloechus* spp.) in dragonfly second intermediate hosts. *Journal of Parasitology*, 93, 593–607.
- Bosma. N.J. (1931) Alaria mustelae sp. nov. a trematode requiring four hosts. Science, 74, 521-522.
- Bosma, N.J. (1934) The life history of the trematode, *Alaria mustelae*, Bosma, 1931. *Transactions of the American Microscopical Society*, 53, 116–153.
- Brandt, B.B. (1936) Parasites of certain North Carolina Salientia. Ecological Monographs, 6, 491-532.
- Bray, R.A., Jones, A. & Andersen, K.I. (1994) Order Pseudophyllidea Carus, 1863. In: Khalil, L.F., A. Jones & R.A. Bray (eds.), *Keys to the cestode parasites of vertebrates*, CAB International, Wallingford, U.K., 205–247.
- Britt, H.G. (1947) Chromosomes of digenetic trematodes. American Naturalist, 81, 276–296.

Brooks, D.R. (1974) Cladistics of frog lung flukes in Nebraska. Proceedings of the Nebraska Academy of Sciences, 84, 7.

Brooks, D.R. (1975a). Systematic studies on the platyhelminth parasites of Nebraska amphibians. M. Sc. Thesis. University of Nebraska, Lincoln.

Brooks, D.R. (1975b) A review of the genus Allassostomoides Stunkard, 1924 (Trematoda: Paramphistomidae) with a

redescription of A. chelydrae (MacCallum 1919) Yamaguti 1958. Journal of Parasitology, 61, 882-885.

- Brooks, D.R. (1976) Parasites of the amphibians of the Great Plains. Part 2. Platyhelminths of amphibians in Nebraska. *Bulletin of the Nebraska State Museum*, 10, 65–92.
- Brooks, D.R. (1977) Evolutionary history of some Plagiorchioid trematodes of anurans. Systematic Zoology, 26, 277–289.
- Brooks, D.R. (1979) New records for amphibian and reptile trematodes. *Proceedings of the Helminthological Society of Washington*, 46, 286–289.
- Brooks, D.R. & Ferrao, A.L. (2005) The historical biogeography of co-evolution: emerging infectious diseases are evolutionary accidents waiting to happen. *Journal of Biogeography*, 32, 1291–1299.
- Brooks, D.R., León-Règagnon, V., McLennan, D.A. & Zelmer, D. (2006a). Ecological fitting as a determinant of the community structure of platyhelminth parasites of anurans. *Ecology*, 87, S76–S85.
- Brooks, D.R. & McLennan, D.A. (2003) *Parascript: Parasites and the language of evolution*. Smithsonian Institution Press, Washington, 448 pp.
- Brooks, D.R., McLennan, D.A., León-Règagnon, V. & Hoberg, E. (2006b) Phylogeny, ecological fitting and lung flukes: helping solve the problem of emerging infectious diseases. *Revista Mexicana de Biodiversidad*, 77, 225–233.
- Buhler, G.A. (1968) Development of the larval stages of *Ophiotaenia gracilis*, a cestode parasite of bullfrogs. *Journal of Colorado-Wyoming Academy of Science*, 6, 59.
- Buhler, G.A. (1970) The post-embryonic development of *Ophiotaenia gracilis* Jones, Cheng and Gillespie, 1958, a cestode parasite of bullfrogs. *Journal of Wildlife Diseases*, 6, 149–151.
- Bursey, C.R. & DeWolf, W.F. II. (1998) Helminths of the frogs, *Rana catesbeiana*, *Rana clamitans*, and *Rana palustris*, from Coshocton County, Ohio. *Ohio Journal of Science*, 98: 28–29.
- Bursey, C.R. & Goldberg, S.R. (2001) *Falcaustra lowei* n. sp. and other helminths from the Tarahumara frog, *Rana tarahumarae* (Anura: Ranidae), from Sonora, Mexico. *Journal of Parasitology*, 87, 340–344.
- Bursey, C.R. & Goldberg, S.R. (2003) Acanthocephalus saurius n. sp. (Acanthocephala: Echinorhynchidae) and other helminths from the lizard Norops limifrons (Sauria: Polychrotidae) from Costa Rica. Journal of Parasitology, 89, 573– 576.
- Bursey, C.R. & Goldberg, S.R. (2005) New species of Oswaldocruzia (Nematoda: Molineoidae), new species of Rhabdias (Nematoda: Rhabdiasidae), and other helminths in Rana cf. forreri (Anura: Ranidae) from Costa Rica. Journal of Parasitology, 91, 600–6005.
- Bursey, C.R. & Goldberg, S.R. (2007) New species of *Hedruris* (Nematoda: Hedruridae), *Anuracanthorhynchus lutzi* (Hamann, 1891) n. comb. and other helminths in *Lithobates warszewitschii* (Anura: Ranidae) from Costa Rica. *Caribbean Journal of Science*, 43, 1–10.
- Bursey, C.R., Goldberg, S.R. & Kraus, F. (2006a) A new species of *Cosmocerca* (Nematoda, Cosmocercidae) and other helminths from *Genyophryne thomsoni* (Anura, Microhylidae) from Papua New Guinea. *Acta Parasitologica*, 51, 213–216.
- Bursey, C.R., Goldberg, S.R. & Vitt, L.V. (2006b) New species of *Oswaldocruzia* (Nematoda: Molineoidae) in *Ameiva festiva* (Squamata: Teiidae) from Nicaragua. *Journal of Parasitology*, 92, 350–352.
- Bush, A.O., Lafferty, K.D., Lotz, J.M. & Shostak, A.W. (1997) Parasitology meets ecology on its own terms: Margolis et al. revised. *Journal of Parasitology*, 83, 575–583.
- Bychowsky, B.E. (1933) Eine neue Gyrodactylus Art aus den Seen Kareliens. Trudy Borodinsk Biologiya Stantsii Karelii, 6, 51–55.
- Bychowsky, B.E. (1937) Ontogenez i filogeneticheskie vzaimootnosheniia ploskikh paraziticheskikh chervei. *Izdatelstvo Akademii Nauk SSSR*, 4, 1353–1383.
- Caballero, E. & Bravo-Hollis, M. (1949) Description d'un nouveau genre de Pleurogeninae (Trematoda: Lecithodendriidae) de grenouilles du Mexique *Langeronia macrocirra* n. g. n. sp. *Annales d'Parasitologie Humainee et Comparee*, 24,193–199.
- Cable, R.M. (1935) *Cercaria kentuckiensis* n.sp., first representative of the *Vivax* group known to occur in the United States. *Journal of Parasitology*, 21, 436.
- Cabrera-Guzmán, E., León-Règagnon, V. & García-Prieto, L. (2007) Helminths of the leopard frog *Rana* cf. *forreri* (Amphibia: Ranidae) in Acapulco, Guerrero, Mexico. *Comparative Parasitology*, 74, 96–107.
- Cain, G.D. & French, J.A. (1975) Effects of parasitism by the lung fluke, *Haematoloechus medioplexus*, on lung fatty acis and sterol composition in the bullfrog, *Rana catesbeiana*. *International Journal for Parasitology*, 5, 159–164.
- Campbell, R.A. (1968) A comparative study of the parasites of certain Salientia from Pocahontas State Park, Virginia. *Virginia Journal of Science*, 19, 13–20.
- Canavan, W.P.N. (1929) Nematode parasites of vertebrates in the Philadelphia Zoological Garden and vicinity. I. *Parasitology*, 21, 63–102.
- Causey, O.R. (1939a) The development of frog filarial larvae, *Foleyella ranae* in *Aedes* and *Culex* mosquitos. *American Journal of Hygiene*, 29, 131–132.
- Causey, O.R. (1939b) Description of three species of frog microfilariae with notes of staining methods. *American Journal* of Hygiene, 30, 117–121.
- Chandler, A.C. (1923) Three new trematodes from Amphiuma means. Proceedings of the U.S. National Museum, 63, 55-

58.

- Chandler, A.C. & Rausch, R. (1947) Study of strigeids from owls in North Central United States. *Transactions of the American Microscopical Society*, 66, 283–292.
- Chen, H.T. (1933) A preliminary report on a survey of animal parasites of Canton China rats. *Lingnan Scientific Journal*, 12, 65–74.
- Cheng, T.C. (1973) Parasitología General. Academic Press, Madrid, 965 pp.
- Choudhury, A. & Dick, T.A. (1998) Patterns and determinants of helminth communities in the Acipenseridae (Actinopterygii: Chondrostei), with special reference to the lake sturgeon, *Acipenser fulvescens*. *Canadian Journal of Zoology*, 76, 330–349.
- Choudhury, A. & León-Règagnon, V. (2005) Molecular phylogenetics and biogeography of *Bunodera* spp. (Trematoda: Allocreadiidae), parasites of percid and gasterosteid fishes. *Canadian Journal of Zoology*, 83, 1540–1546.
- Christensen, B.M. (1981) A taxonomic review of the genus *Loxogenoides* (Digenea: Lecithodendriidae) with a description of *Loxogenoides loborchis* sp. n. from *Rana catesbeiana* Shaw in western Kentucky. *Proceedings of the Helminthological Society of Washington*, 48, 65–70.
- Christian, F.A. (1971) *Pseudosonsinotrema catesbianae* sp. n. (Trematoda: Pleurogenidae) from the bullfrog, *Rana catesbeiana* Shaw. *Proceedings of the Helminthological Society of Washington*, 38, 37–39.
- Clark, R.T. & Longest, W.D. (1969) The distribution and occurrence of frog lung flukes in Lafayette, Pontotoc, and Panola Counties, Mississippi. *Journal of the Mississippi Academy of Sciences*, 15, 63.
- Corkum, K.C. (1966) Sparganosis in some vertebrates of Louisiana and observations on a human infection. *Journal of Parasitology*, 52, 444–448.
- Cort, W.W. (1912) North American frog bladder flukes. *Transactions of the American Microscopical Society*, 31, 151–166.
- Cort, W.W. (1913) Notes on the trematode genus *Clinostomum*. *Transactions of the American Microscopical Society*, 32, 169–182.
- Cort, W.W. (1914) Larval trematodes from North American freshwater snails. Preliminary report. *Journal of Parasitol*ogy, 1, 65–84.
- Cort, W.W. (1915) North American frog lung flukes. Transactions of the American Microscopical Society, 34, 203-240.
- Cort, W.W. (1919) A new distome from Rana aurora. University of California Publications in Zoology, 19, 283–289.
- Cort, W.W. (1926) Megalodiscus Chandler should sink in favor of Diplodiscus Dies. Journal of Parasitology, 12, 180.
- Coy-Otero, A. & Barus, V. (1979) Nematodes parasitizing Cuban reptiles. *Acta Scientarum Naturalium Academiae Scientiarum Bohemicae Borneo*, 13, 1–43.
- Coy-Otero, A. & Martinez, J. (1987) Nuevo hallazgo en Cuba de larvas de *Eustrongyloides* Jaegerskiöld 1909 (Nematoda: Dyoctophrynidae) en la rana toro (*Rana catesbeiana*). *Miscelanea Zoológica de la Academia de Ciencias de Cuba*, 17, 3.
- Coy-Otero, A. & Ventosa, M.L. (1984) Nemátodos parásitos de anfibios cubanos. Poeyana, 269, 1-20.
- Crans, W.J. (1969) Preliminary observations of frog filariasis in New Jersey. *Bulletin of the Wildlife Disease Association*, 5, 342–347.
- Crawshaw, G.J. (1997). Disease in Canadian amphibian populations. Herpetological Conservation, 1, 258–270.
- de Chambrier, A., Coquille, S.C. & Brooks, D.R. (2006) *Ophiotaenia bonneti* sp. n. (Eucestoda: Proteocephalidea), a parasite of *Rana vaillanti* (Anura: Ranidae) in Costa Rica. *Folia Parasitologica*, 53, 125–133.
- de Chambrier, A., Zehnder, M.P., Vaucher, C. & Mariaux, J. (2004) The evolution of the Proteocephalidea (Platyhelminthes, Eucestoda) based on an enlarged molecular phylogeny, with comments on their uterine development. *Systematic Parasitology*, 57, 159–171.
- Dronen, N.O. (1977) Studies on the population structure of two species of *Haematoloechus* Looss, 1899 (Digenea: Plagiorchiidae) in raniid frogs in New Mexico. *Proceedings of the Helminthological Society of Washington*, 44, 68–72.
- Durette-Desset, M-C., Alves Dos Anjos, L. & Vrcibradic, D. (2006) Three new species of the genus Oswaldocruzia Travassos, 1917 (Nematoda: Trichostrongylina, Molineoidea) parasites of Enyalius spp. (Iguanidae) from Brazil. Parasite, 13, 115–125.
- Dzikowski, R., Levy, M.G., Poore, M.F., Flowers, J.R. & Paperna, I. (2004) *Clinostomum complanatum* and *Clinostomum marginatum* (Rudolphi, 1819) (Digenea: Clinostomidae) are separate species based on differences in ribosomal DNA. *Journal of Parasitology*, 90, 413–414.
- Esch, G.W. & Kocan, R. (1966) *Teloporia* (Trematoda: Pronocephalidae) from an amphibian. *Journal of Parasitology*, 52, 497.
- Espinoza-Jiménez, A., García-Prieto, L., Osoro-Sarabia, D. & León-Règagnon, V. (2007) Checklist of the helminth parasites of *Bufo marinus* in Mexico. *Journal Parasitology*, 93, 937–944.
- Esslinger, J.H. (1986) Redescription of *Foleyellides striatus* (Ochoterena & Caballero, 1932) (Nematoda: Filarioidea) from a Mexican frog, *Rana montezumae*, with reinstatement of the genus *Foleyellides* Caballero, 1935. *Proceedings of the Helminthological Society of Washington*, 53, 218–223.
- Fantham, H.B. & Porter, A. (1948) The parasitic fauna of vertebrates in certain Canadian freshwaters with some remarks on their ecology, structure, and importance. *Proceedings of the Zoological Society of London*, 117, 609–649.

Faust, E.C. (1918) Studies on Illinois cercariae. Journal of Parasitology, 4, 93–110.

- Freitas, J.F.T. (1941) Sobre alguns trematódeos de ras. Revista Brasileira de Biologia, 1, 122-123.
- Freitas, J.F.T. & Lent, H. (1941) Contribuicao ao conhecimento da sub-familia Kathlaniinae Lane, 1914 (Nematoda: Subuluroidea). *Arquivos Zoologicos do Estado de Sao Paulo*, 3, 13–41.
- Freze, V.I. & Rysavy, B. (1976) Cestodes of the suborder Proteocephalata Spassky, 1957 (Cestoda-Pseudophyllidea) from Cuba and description of a new species *Ophiotaenia habanensis* sp. n. *Folia Parasitologica*, 23, 97–104.
- Fried, B. & Bradford, J.D. (1997) In vitro excystation of metacercarial cyst of *Echinostoma trivolvis* from *Rana* species tadpoles. *Korean Journal of Parasitology*, 35, 75–77.
- Frölich, J.A. (1789) Beschreibungen einiger neuer Eingeweidwürmer. Naturforscher Halle, 24, 101–162.
- Frost, D.R., Grant, T., Faivovich, J., Bain, R., Haas, A., Haddad, C.F.B., de Sá, R.O., Donnellan, S.C., Raxworthy, C.J., Wilkinson, M., Channing, A., Campbell, J.A., Blotto, B.L., Moler, P., Drewes, R.C., Nussbaum, R.A., Lynch, J.D., Green, D. & Wheeler, W.C. (2006) The amphibian tree of life. *Bulletin of the American Museum of Natural History*, 297, 370 pp.
- Führmann, O. (1928) Zweite klasse des cladus Plathelminthes: Trematoda. Handb Zoologischer, 2, 1–140.
- Fukui, T. (1929) Studies on Japanese amphistomatous parasites, with revision of the group. *Japanese Journal of Zoology*, 2, 91–102.
- Fukui, T. (1933) Teloporia (Tremat.) = Opisthoporus. Zoologischer Anzeiger, 103, 332–333.
- Garner, T.W.J., Perkins, M.W., Govindarajulu, P., Seglie, D., Walker, S., Cunningham, A.A. & Fisher, M.C. (2006) The emerging amphibian pathogen *Batrachochytrium dendrobatidis* globally infects introduced populations of the North American bullfrog, *Rana catesbeiana*. *Biology Letters*, 2, 455–459.
- Gastaldi, B. (1854) Cenni sopra alcuni nuovi elminti della *Rana esculenta*, con nuove osservazione sul *Codonocephalus mutabilis* Dies., Torino, 14 pp.
- Global Invasive Species Database (2005) *Lithobates catesbeianus*. Available from: http://www.issg.org/database/species/ ecology.asp?si=19&fr=1&sts=sss [Accessed July 2009].
- Goeze, J.A. (1782) Versuch einer Naturgeschichte der Eingeweidewürmer thierischer Körper. XI + 471 pp. Blankenburg.
- Goldberg, S.R. & Bursey, C.R. (2002a) Helminths of the bullfrog, *Rana catesbeiana* (Ranidae), in California with revisions to the California anuran helminth list. *Bulletin* (*Southern California Academy of Sciences*). Journal on line.
- Goldberg, S.R. & Bursey, C.R. (2002b) Helminths of 10 Species of Anurans from Honshu Island, Japan. *Comparative Parasitology*, 69, 162–176.
- Goldberg, S.R. & Bursey, C.R. (2007) Helminths of two species of frogs, *Lithobates taylori* and *Lithobates vaillanti* (Ranidae), from Costa Rica. *Caribbean Journal of Science*, 43, 65–72.
- Goldberg, S.R. & Bursey, C.R. (2008) Helminths from 10 species of Brachycephalid frogs (Anura: Brachycephalidae) from Costa Rica. *Comparative Parasitology*, 75, 255–262.
- Goldberg, S.R., Bursey, C.R. & Cheam, H. (1998) Helminths of two native frog species (*Rana chiricahuensis*, *Rana yava-paiensis*) and one introduced frog species (*Rana catesbeiana*) (Ranidae) from Arizona. *Journal of Parasitology*, 84, 175–177.
- Goldberg, S.R., Bursey, C.R., McKinnell, R.G. & Tan, I.S. (2001) Helminths of northern leopard frogs, *Rana pipiens* (Ranidae) from North Dakota and South Dakota. *Western North American Naturalist*, 61, 248–251.
- Goldberg S.R., Bursey, C.R. & Walser, C.M. (2000) Intestinal helminths of seven species of agamid lizards from Australia. *Comparative Parasitology*, 67, 109–114.
- Goldberg, S.R., Bursey, C.R. & Wong, C. (2002) Helminths of the western chorus frog from Eastern Alberta, Canada. *Northwest Science*, 76, 77–79.
- Goodchild, C.G. (1945) Additional observations on the life history of *Gorgodera amplicava* Looss, 1899. *Journal of Parasitology*, 31, 22–23.
- Goodchild, C.G. (1948) Additional observations on the bionomics and life history of *Gorgodera amplicava* Looss, 1899 (Trematoda: Gorgoderidae). *Journal of Parasitology*, 34, 407–427.
- Goodchild, C.G. (1950) Establishment and pathology of gorgoderid infections in anuran kidneys. *Journal of Parasitology*, 36, 439–446.
- Goodchild, C.G. (1954) Survival of gorgoderine trematodes into challenging habitats. *Experimental Parasitology*, 40, 591–602.
- Goodchild, C.G. (1955) Transplantation of gorgoderine trematodes into challenging habitats. *Experimental Parasitology*, 4, 351–60.
- Guberlet, J.E. (1919) A new bladder fluke from the frog. *Proceedings of the Seventeenth Annual Meeting of the American Society of Zoologists (Abstracts), Anat. Record*, 17, 331.
- Guberlet, J.E. (1920) A new bladder fluke from the frog. *Transactions of the American Microscopical Society*, 39, 142–148.
- Hannum, C.A. (1925) A new species of cestode, *Ophiotaenia magna* n. sp. from the frog. *Transactions of the American Microscopical Society*, 44, 148–155.
- Harwood, P.D. (1930) A new species of *Oxysomatium* (Nematoda) with some remarks on the genera *Oxysomatium* and *Aplectana*, and observations on the life history. *Journal of Parasitology*, 17, 61–73.

- Harwood, P.D. (1932) The helminths parasitic in the Amphibia and Reptilia of Houston, Texas and vicinity. *Proceedings* of the US National Museum, 81, 1–71.
- Hasegawa, H. (2006) First record of *Falcaustra catesbeianae* Walton, 1929 (Nematoda, Cosmocercoidea, Kathlaniidae) from the Bullfrog, *Rana catesbeiana*, in Japan. *Biogeography*, 8, 1–5.
- Hasegawa, H. & Asakawa, M. (2004) Parasitic nematodes recorded from amphibians and reptiles in Japan. *Current Herpetology*, 23, 27–35.
- Hedrick, L.R. (1935) Taxonomy of the nematode genus *Spiroxys* (Family Spiruridae). *Journal of Parasitology*, 21, 397–409.
- Hoffman, G.L. & Dunbar, C.E. (1963) Studies on *Neogogatea kentuckiensis* (Cable, 1935) n. comb. (Cyathocotylidae). *Journal of Parasitology*, 49, 737–744.
- Holl, F.J. (1928) Two new nematode parasites. Journal of the Elisha Mitchell Science Society, 43, 184–186.
- Hollis, P.D. (1972) A survey of parasites of the bullfrog, *Rana catesbeiana* Shaw, in Central East Texas. *Southwestern Naturalist*, 17, 198–200.
- Hunt, G.W. (1952) The life history of *Gorgodera vivata* n. sp. (Trematoda: Gorgoderidae). *Dissertation Abstract International*, 12, 233–234.
- Hunter, G.W. III. (1930) *Diplodiscus intermedius* sp. nov., from *Rana catesbeiana* Shaw. *Journal of Parasitology*, 17, 74–79.
- Ingles, I.G. (1936) Worm parasites of California amphibia. *Transactions of the American Microscopical Society*, 55, 73–92.
- Irwin, M.S. (1929) A new lung fluke from *Rana clamitans* Latreille. *Transactions of the American Microscopical Society*, 48, 74–79.
- Jacobs, B. & Morrison, E.O. (1966) Ectopic lung fluke. Southwestern Naturalist, 11, 412.
- Jewell, M.E. (1916) Cylindrotaenia americana nov. spec. from the cricket frog. Journal of Parasitology, 2, 181–193.
- Jinks, J.L. & Johnston, J.C. Jr. (1971) Trematodes of *Rana catesbeiana* from three strip-mine lakes in southeast Kansas. *Transactions of the Kansas Academy of Sciences*, 73, 519–520.
- Johnson, A.D. (1968) Life history of *Alaria marcianae* (La Rue, 1917) Walton, 1949 (Trematoda: Diplostomatidae). *Journal of Parasitology*, 54, 324–332.
- Johnson, A.D. (1979) Morphology and life history of *Alaria mustelae* Bosma 1931 (Trematoda: Diplostomatidae) from Minnesota mustelids. *Journal of Parasitology*, 65, 154–60.
- Johnson, P.T.J., Lunde, K.B., Ritchie, E.G. & Launer, A.E. (1999) The effect of trematode infection on amphibian limb development and survivorship. *Science*, 284, 802-804.
- Johnson, P.T.J., Lunde, K.B., Thurman, E.M., Ritchie, E.G., Wray, S.N., Sutherland, D.R., Kapfer, J.M., Frest, T.J., Bowerman, J. & Blaustein, A.R. (2002) Parasite (*Riberoia ondatrae*) infection linked to amphibian malformations in the western United States. *Ecological Monographs*, 72, 151–168.
- Jones, A.W., Cheng, T.C. & Gillespie, R.F. (1958) *Ophiotaenia gracilis* n. sp. a proteocephalid cestode from a frog. *Journal of Tennessee Academy of Sciences*, 33, 84–88.
- Jones, A., Bray, R.A. & Gibson, D.I. (2005) Keys to the Trematoda. Vol. 2. CABI Publishing and The Natural History Museum, London.
- Karmanova, E.M. (1961) The first report of *Dioctophyme renale* in fish. In. E. USSR. Akademi Nauka, SSSR, 11, 118–121.
- Kaw, B.L. (1945) On the present status of the genus *Loxogenes*. *Proceedings of the Indian Academy of Sciences*, 21, 342–343.
- Kaw, B.L. (1950) Studies in helminthology: helminth parasites of Kashmir. Part I. Trematoda. *Indian Journal of Helminthology*, 2, 6–126.
- Kennedy, M.J. (1981) A revision of species of the genus *Haematoloechus* Looss, 1899 (Trematoda: Haematoloechidae) from Canada and the United States. *Canadian Journal of Zoology*, 59, 1836–1846.
- Kim, K-H., Rim, H-J. & Yoon, I-B. (1992) Trematodes of the genus *Haematoloechus* (Digenea: Plagiorchiidae) from frogs in Korea. *The Korean Journal of Parasitology*, 30, 245–253.
- Knight, M.T., Barbay, C.J. & Morrison, E. (1965) Incidence and infection by lung-fluke (*Haematoloechus*) of the bullfrog, *Rana catesbiana*, in Jefferson County, Texas. *Southwestern Naturalist*, 10, 141–142.
- Kotcher, E. (1941) Studies on the development of frog filariae. American Journal of Hygiene, 34, 36-65.
- Krissinger, W.A. & Mehra, K.N. (1968) Studies on the biology of *Proterometra albacauda* Anderson and Anderson, 1967, an Azygiid trematode. *ASB Bulletin*, 15, 43.
- Krull, W.H. (1930) The life history of two North American frog lung flukes. Journal of Parasitology, 16, 207–212.
- Krull, W.H. (1932) Studies on the life history of *Pneumobites longiplexus* (Stafford). *Zoologische Anzeiger*, 99, 231–239. Krull, W.H. (1933) *Loxogenes bicolor*, a new pigmented fluke from the frog, *Rana clamitans. Transactions of the Ameri-*
- can Microscopical Society, 52, 47–50.
 Krull WH (1935) Studies on the life history of a free bladder fluke. Correctory annullary Looss, 1800. Banars of the
- Krull, W.H. (1935) Studies on the life history of a frog bladder fluke, *Gorgodera amplicava* Looss, 1899. *Papers of the Michigan Academy of Science, Arts, and Letters*, 20, 697–710.
- Krull, W.H. & Price, H.F. (1932) Studies on the life history of Diplodiscus temperatus Stafford from the frog. Occasional

Papers of the Museum of Zoology, University of Michigan, 237, 1–37.

- Kung, C.C. (1948) On new some species of spirurids from terrestrial vertebrates, with notes on *Habronema mansoni*, *Physaloptera paradoxa*, and *Hartertia zuluensis*. *Journal of Helminthology*, 22, 141–164.
- Kuntz, R.E. & Self, J.T. (1944) An ecological study of the Metazoan parasites of the Salientia of Comanche County, Oklahoma. *Proceedings of the Oklahoma Academy of Sciences*, 24, 35–38.
- Kuzmin, Y., Tkach, V.V. & Snyder, S.D. (2003) The nematode genus *Rhabdias* (Nematoda: Rhabdiasidae) from amphibians and reptiles of the Nearctic. *Comparative Parasitology*, 70, 101–114.
- Lamothe-Argumedo, R. & García-Prieto, L. (1988) *Helmintiasis del hombre en México. Tratamiento y Profilaxis*. AGT Editores, Mexico City, 139 pp.
- La Rue, G.R. (1917) Two new larval trematodes from *Thamnophis marciana* and *Thamnophis eques*. Ocassional Papers of the Museum of Zoology, University of Michigan, 35, 1–12.
- Lang, B.Z. (1968) The life cycle of *Cephalogonimus americanus* Stafford, 1902 (Trematoda: Cephalogonimidae). *Journal of Parasitology*, 54, 945–949.
- Lank, D.R. (1971) Parasites of the bullfrog in Indiana. Proceedings of the Indiana Academy of Science, 81, 359-364.
- Lehmann, D.L. (1965) Intestinal parasites of northwestern amphibians. *Yearbook of the American Philosophical Society*, 1965, 284–285.
- Leidy, J. (1851) Contributions to Helminthology. *Proceedings of the Academy of Natural Sciences of Philadelphia*, 5, 249–351.
- Leidy, J. (1856) A synopsis of Entozoa and some of the other ecto-congeners observed by the author. *Proceedings of the Academy of Natural Sciences of Philadelphia*, 8, 42–58.
- Leigh, W.H. (1937a) The life-cycle of a trematode of frogs. Science, 86, 423.
- Leigh, W.H. (1937b) The life-cycle of a trematode of frogs. Journal of Parasitology, 23, 563.
- Leigh, W.H. (1946) Experimental studies on the life cycle of *Glypthelmins quieta* (Stafford, 1900), a trematode of frogs. *American Midland Naturalist*, 35, 460–483.
- Leigh, W.H. & Van Cleave, H. (1945) Metamorphosis of the frog host as a factor in cercarial penetration by *Glypthelmins quieta*. *Journal of Parasitology*, 31, 205–209.
- Lemke, L.B., Dronen, N., Fox, J.G. & Nambiar, P.R. (2008) Infestation of wild-caught American bullfrogs (*Rana catesbe-iana*) by multiple species of metazoan parasites. *Journal of the American Association for Laboratory Animal Science*, 47, 42–46.
- León-Règagnon, V. (2003) Incorporating morphological and molecular data in biodiversity inventories. Parasites of leopard frogs. *Journal of Parasitology*, 89, S141-S148.
- León-Règagnon, V. & Brooks, D.R. (2003) Molecular phylogeny of *Haematoloechus* Looss, 1899 (Digenea: Plagiorchiidae), with emphasis on North American species. *Journal of Parasitology*, 89, 1206–1211.
- León-Règagnon, V., Brooks, D.R. & Perez-Ponce de León, G. (1999) Differentiation of Mexican species of *Haema-toloechus* Looss, 1899 (Digenea: Plagiorchiformes): Molecular and morphological evidence. *Journal of Parasitology*, 85, 935–946.
- León-Règagnon, V., Gillén-Hernández, S. & Arizmendi-Espinoza, M.A. (2005) Intraspecific variation of *Haematoloechus floedae* Harwood, 1932 (Digenea: Plagiorchidae) from *Rana* spp. from North and Central America. *Journal of Parasitology*, 91, 915–921.
- León-Règagnon, V. & Paredes-Calderón, L. (2002) *Haematoloechus danbrooksi* n. sp. (Digenea: Plagiorchioidea) from *Rana vaillanti* from Los Tuxtlas, Veracruz, Mexico. *Journal of Parasitology*, 88, 1215–1221.
- Linnaeus, C. (1758) Systema Naturae per regna tria naturae secundum classes, ordines, genera, species, cum characteribus, differentitiis, synonymis, locis. Editio decimal, reformata, I, 823 pp. Holmiae.
- Loftin, H. (1960) An annotated checklist of trematodes and cestodes and their vertebrate hosts from northwest Florida. *Quarterly Journal of the Florida Academy of Science*, 23, 302–324.
- Looss, A. (1899a) Weitere beitrage zur kenntnis der trematodenfauna Aegyptens, zugleich versuch einernaturlichen gliederung des genus *Distomum* retzius. *Zoologischer Jahrbuecher Systematik*, 12, 521–784.
- Looss, A. (1899b) Weitere Beitrage zur Kenntniss der Trematoden-Fauna Aegyptens. Zoologischer Jahrbuecher Systematik, 12, 605–607.
- Looss, A. (1905) Ueber neve und bekkante Trematoden aus Seechildtröten. Zoologische Jahrbücher Abtheilung für Systematik, 16, 411–894.
- MacCallum, G.A. (1921) Studies in helminthology Part 1. Trematodes. Zoopathologica, 1, 137-284.
- Mace, T.F. & Anderson, R.C. (1975) Development of the giant kidney worm, *Dioctophyma renale* (Goeze, 1782) (Nematoda: Dioctophymatoidea). *Canadian Journal of Zoology*, 53, 1552–1568.
- Macy, R.W. (1964) A new species of trematode, *Pycnoporus remsesi* (Lecithodendriidae) from Egypt and notes on *P. acetabulatus* Looss, 1899. *Proceedings of the Helminthological Society of Washington*, 31, 292–297.
- Manter, H.W. (1938) A collection of trematodes from Florida amphibia. Transactions of the American Microscopical Society, 57, 26–38.

Martínez, J., Coy, A. & Ventosa, L. (1982) Helmintos de *Rana catesbeiana* Shaw (Ranidae) en Cuba. *Poeyana*, 243, 2–10. Martínez-Salazar, E. (2004) *Estudio taxonómico de algunas poblaciones del género Langeronia Caballero y Bravo*, 1949

(*Trematoda: Lecithodendriidae*) en México. M.Sc. Thesis, Facultad de Ciencias, Universidad Nacional Autónoma de México, Mexico City, 92 pp.

- Martínez-Salazar, E.A. (2006) A new rhabdiasid species from *Norops megapholidotus* (Sauria: Polychrotidae) from Mexico. *Journal of Parasitology*, 92, 1325–1329.
- Mata-López, R., León-Règagnon, V. & Brooks, D.R. (2005) Species of *Gorgoderina* (Digenea: Gorgoderidae) in *Rana vaillanti* and *Rana* cf. *forreri* (Anura: Ranidae) from Guanacaste, Costa Rica, including a description of a new species. Journal of Parasitology, 91, 403–410.
- Mayberry, L.F., Canaris, A.G., Bristol, J.R. & Gardner, S. L. (2000) Bibliography of parasites and vertebrate hosts in Arizona, New Mexico and Texas (1893 1984). Electronic publication of the Harold W. Manter Laboratory of Parasitology, University of Nebraska, Lincoln, Nebraka, 100 pp.
- McAllister, C.T., Bursey, C.R. & Trauth, S.E. (2008) New host and geographic distribution records for some endoparasites (Myxosporea, Trematoda, Cestoidea, Nematoda) of amphibians and reptiles from Arkansas and Texas, U.S.A. *Comparative Parasitology*, 75, 241–254.
- MacCallum, G.A. (1917) Some new forms of parasitic worms. Zoopathologica, 1, 42–75.
- MacCallum, G.A. (1919) Notes on the genus *Telorchis* and other trematodes. *Zoopathologica*, 1, 88–89.
- McAlpine, D.F. (1996) Acanthocephala parasitic in North American amphibians: a review with new records. *Alytes*, 14, 115–121.
- McAlpine, D.F. (1997a) Systematics and ecology of frog helminths in New Brunswick, Canada. *Dissertation Abstracts International Part B: Science and Engineering*, 58, 2902.
- McAlpine, D.F. (1997b) Helminth communities in bullfrogs (*Rana catesbeiana*), green frogs (*Rana clamitans*), and leopard frogs (*Rana pipiens*) from New Brunswick, Canada. *Canadian Journal of Zoology*, 75, 1883–1890.
- McAlpine, D.F. (2006) *Halipegus occidualis* Krull, 1935 and *Halipegus eccentricus* Thomas, 1939 (Digenea, Hemiuridae): proposed conservation of the specific names. *Bulletin of Zoological Nomenclature*, 63, 12–16.
- McAlpine, D.F. & Burt, M.D.B. (1998a) Helminths of Bullfrogs, *Rana catesbeiana*, Green Frogs, *R. clamitans* and Leopard Frogs, *R. pipiens*, in New Brunswick. *Canadian Field- Naturalist*, 112, 50–68.
- McAlpine, D.F. & Burt, M.D.B. (1998b) Taxonomic status of *Halipegus* spp. (Digenea: Derogenidae) parasitic in the mouth and Eustachian tubes of North American and Mexican amphibians. *Journal of the Helminthological Society of Washington*, 65, 10–15.
- Meggitt, F.J. (1925) On the life history of an amphibian tapeworm, *Diphyllobothrium ranarum*. *Annals Magazine of Natural History*, 16, 654–655.
- Merritt, S.V. & Pratt, I. (1964) The life history of *Neoechinorhynchus rutili* and its development in the intermediate host (Acanthocephala: Neoechinorhynchidae). *Journal of Parasitology*, 50, 394–400.
- Miller, E.L. (1930) Studies on *Glypthelmins quieta* Stafford. Journal of Parasitology, 16, 237–243.
- Miller, G.C., Harkema, R. & Harris, A. (1965a) Life history notes on *Strigea elegans* Chandler and Rausch, 1947 (Trematoda: Strigeidae). *Journal of Parasitology*, 51, 22.
- Miller, G.C., Harkema, R. & Harris, A. (1965b) Notes on the life history of *Strigea elegans* Chandler and Rausch, 1947 (Trematoda: Strigeidae). *Journal of Parasitology*, 51, 894–895.
- Miller, G.C., Harkema, R. & Harris, A. (1965c) Life history notes on *Strigea elegans* Chandler and Rausch, 1947 (Trematoda: Strigeidae). *Journal of Elisha Mitchell Science Society*, 81, 86–87.
- Miyazaki, I. (1991) An illustrated book of helminthic zoonoses. International Medical Foundation of Japan, Fukuoka, Japan, 494 pp.
- Modzelewski, E. & Culley, D.D. (1974) Ocurrence of the nematode *Eustrongylides wenrichi* in laboratory reared *Rana* catesbeiana. Copeia, 1974, 1000–1001.
- Moravec, F. (1998) Nematodes of freshwater fishes of the Neotropical Region. Academia, Praha. 464 pp.
- Moravec, F. & Kaiser, H. (1995) Helminth parasites from West Indian frogs, with descriptions of two new species. *Caribbean Journal of Science*, 31, 252–268.
- Morgan, B.B. (1941) Additional notes on North American Physalopterinae (Nematoda). *Proceedings of the Helminthological Society of Washington*, 8, 63–64.
- Morrison, E.O. (1966) Crayfish, posible secondary intermediate host for lung flukes (*Haematoloechus*) of bullfrogs in Jefferson County, Texas. *Yearbook of the American Philosphical Society*, 1966, 361-362.
- Morrison, E.O. (1967) *Rhabdias ranae* infections in three species of Oklahoma Anura. *Southwestern Naturalist*, 12, 335–336.
- Mueller, J.F. (1935) A Diphyllobothrium from cats and dogs in the Syracuse region. Journal of Parasitology, 22, 471–478.

Mueller, J.F. (1974) The biology of Spirometra. Journal of Parasitology, 60, 3–14.

- Müller, O.F. (1780) Unterbrochene Bemühungen bei den intestinalwürmern. Schriften Berlinische Gesellschaft Naturforschender Freunde, 1, 202–218.
- Muzzall, P.M. (1991) Helminth infracommunities in frogs *Rana catesbeiana* and *Rana clamitans* from Turkey Marsh, Michigan. *Journal of Parasitology*, 77, 366–371.
- Muzzall, P.M. (2005) Parasites of amphibians and reptiles from Michigan. A review of the literature 1916–2003. *State of Michigan Department of Natural Resources, Fisheries Research Report*, 2077, 30 pp.

- Muzzall, P.M. & Baker, M.R. (1987) First report of *Hedruris siredonis* (Nematoda: Hedruridae) from North American frogs. *Proceedings of the Helminthological Society of Washington*, 54, 276–277.
- Muzzall, P.M., Gillilland, M.G.III, Summer, C.S. & Mehne, C.J. (2001) Helminth communities of green frogs *Rana clamitans* Latreille, from Southwestern Michigan. *Journal of Parasitology*, 87, 962–968.
- Myer, D.G. (1960) On the life history of *Mesostephanus kentuckiensis* (Cable, 1935) n. comb. (Trematoda: Cyathocotylidae). *Journal of Parasitology*, 46, 819–832.
- Myers, B.J. & Kuntz, R.E. (1969) Nematodes of fishes, amphibians, and reptiles taken by U.S. Naval Medical Research Unit No. 2 expedition to North Borneo (Malaysia). *Journal of the Fishery Research Board of Canada*, 26, 793–797.
- Nagasawa, K. & Egusa, S. (1981) Acanthocephalus lucidis Van Claeve (Acanthocephala: Echinorhynchidae) from cultured rainbow trout Salmo gairdneri Richardson. Bulletin of the Japanese Society of Scientific Fisheries, 47, 1153– 1156.
- Najarian, H.H. (1955) Trematodes parasitic in the Salientia in the vicinity of Ann Arbour, Michigan. *American Midland Naturalist*, 53, 195–197.
- Nicol, J.T., Demaree, R. Jr. & Wootton, D.M. (1985) Levinseniella (Monarrhenos) ophidea sp. n. (Trematoda: Microphallidae) from the western garter snake, *Thamnophis elegans* and the bullfrog, *Rana catesbeiana*. Proceedings of the Helminthological Society of Washington, 52, 180-183.
- Nickerson, W.S. (1900) Note on Distomum arcanum (n. sp.) in American frogs. American Naturalist, 34, 811-815.
- Nickol, B.B. (1972) *Fessisentis*, a genus of acanthocephalans parasitic in North American poikilotherms. *Journal of Parasitology*, 58, 282–289.
- O'Grady, R.T. (1987) Phylogenetic systematics and the evolutionary history of some intestinal flatworms parasites (Trematoda: Digenea: Plagiorchioiidea) of anurans. Ph. D. Thesis, University of British Columbia, Vancouver, B. C., Canada. 210 pp.
- Ochi, S. (1930) Über die Entwicklungsgeschichte von Mesocoelium brevicaecum n. sp. Okayama-Igakkai-Zasshi, 42, 388-402.
- Odening, K. (1960) Plagiorchiidae III. (Haematoloechinae) und Omphalometrinae. *In*: Mertens, R. & Hennig, W. (Eds) *Das Tierreich. Eine Zusammenstellung und Kennzeichnung der rezenten Tierformen*, Walter de Gruyter & Co., Berlin, Germany, pp. 75.
- Odening, K. (1968) Einige Trematoden aus Fröschen und Schildkröten in Vientnam und Kuba. Zoologischer Anzeiger, 181, 289–302.
- Odlaug, T.O. (1936) Notes on the development of *Gorgodera amplicava* in the final host. *Journal of Parasitology*, 22, 535.
- Odlaug, T.O. (1937) Notes on the development of Gorgodera amplicava in the final host. Biological Bulletin, 72, 80-87.

Odlaug, T.O. (1954) Parasites of some Ohio amphibia. *Ohio Journal of Science*, 54, 126–128.

- Osler, C.P. (1931) A new cestode from Rana clamitans Latr. Journal of Parasitology, 17, 183–186.
- Otsuru, M. (1977) Angiostrongylus cantonensis. In: Sasa (Ed.), Animals of Medical Importance in the Nasei Islands in Japan. Shinjyuku-shobo, Tokyo, pp. 343–374.
- Owen, R. (1836) Anatomical description of two species of Entozoa from the stomach of a tiger (*Felis trigris* Linn.) one of which forms a new genus of Nematoidea, *Gnathostoma. Proceedings of the Zoological Society of London*, 4, 123–126.
- Paetow, L., Cone, D.K., Huyse, T., McLaughlin, D. & Marcogliese, D. (2009) Morphology and molecular taxonomy of *Gyrodactylus jennyae* n. sp. (Monogenea) from tadpoles of captive *Rana catesbeiana* Shaw (Anura), with a review of the species of *Gyrodactylus* Nordmann, 1832 parasitising amphibians. *Systematic Parasitology*, 73, 219–227.
- Pande, B.P. (1937) On the morphology and systematic position of a new bladder fluke from an Indian frog. *Annals of the Magazine Natural History*, 20, 250–256.
- Panesar, T.S. & Beaver, P.C. (1979) Morphology of the advanced-stage larva of *Eustrongylides wenrichi* Canavan 1929, occurring encapsulated in the tissues of *Amphiuma* in Louisiana. *Journal of Parasitology*, 65, 96–104.
- Paredes-León, R., García-Prieto, L., Guzmán-Cornejo, C., León-Règagnon, V. & Pérez, T.M. (2008) Metazoan parasites of Mexican amphibians and reptiles. *Zootaxa*, 1904, 1–166.
- Parker, M.V. (1941) The trematode parasites from a collection of amphibians and reptiles. *Journal of the Tennessee Academy of Sciences*, 16, 27–45.
- Pérez-Ponce de León, G., García-Prieto, L. & Mendoza-Garfias, B. (2007) Trematode parasites (Platyhelminthes) of wildlife vertebrates in Mexico. *Zootaxa*, 1534, 1–247.

Pérez-Vigueras, I. (1938) Notas sobre algunos nematodes parasitos nuevos de Cuba. Livro Jubilar de Travassos, 501–508.

Pérez-Vigueras, I. (1942) Notas helmintológicas. Revista de la Universidad de la Habana, 40-41-42, 193-213.

- Prudhoe, S. & Bray, R. (1982) Plathelminth parasites of the Amphibia. British Museum (Natural History) Oxford University Press, New York, 135 pp.
- Pryor, G.S. & Bjorndal, K.A. (2005) Effects of the nematode *Gyrinicola batrachiensis* on development, gut morphology, and fermentation in bullfrog tadpoles (*Rana catesbeiana*): a novel mutualism. *Journal of Experimental Zoology*, 303, 704–712.
- Pryor, G.S. & Greiner, E.C. (2004) Expanded geographical range, new host accounts, and observations of the nematode

Gyrinicola batrachiensis (Oxyuroidea: Pharyngodonidae) in tadpoles. Journal of Parasitology, 90, 189–191.

- Ralph, P.H. (1938) Cercaria concavocorpa Sizemore becomes Tetrapapillatrema, a new Telorchid-like genus of Plagiorchioidea Dollfus. Transactions of the American Microscopical Society, 57, 376–382.
- Rankin, J.S. (1937) An ecological study of parasites of some North Carolina salamanders. *Ecological Monographs*, 7, 169–269.
- Rankin, J.S. (1938) Studies on the trematode genus *Brachycoelium* Duj. I. Variation in specific characters with reference to the validity of the described species. *Transactions of the American Microscopical Society*, 57, 358–375.
- Rankin, J.S. (1944a) A review of the trematode genus *Glypthelmins* Stafford, 1905, with an account of the life cycle of *G. quieta* (Stafford, 1900) Stafford, 1905. *Transactions of the American Microscopical Society*, 63, 30–43.
- Rankin, J.S. (1944b) A review of the trematode genus *Halipegus* Looss, 1899, with an account of the life history of *H. amhertensis* n. sp. *Transactions of the American Microscopical Society*, 63, 149–164.
- Rankin, J.S. (1945) An ecological study of the helminth parasites of amphibians and reptiles of western Massachusetts and vicinity. *Journal of Parasitology*, 31, 142–150.
- Rao, R. (1977) On a new species of *Pleurogenoides* Travassos, 1971 (Lecithodendriidae Odhner, 1910) and *P. sitapurii*, Srivastava, 1934 from frogs in Hyderabad. *Rivista di Parassitologia*, 38, 23–29.
- Rau, M.E., Doyle, J. & Gordon, D. (1978) Les parasites des animaux sauvages du Quebec. 2. Les parasites des grenouilles et des serpents de la region del l'Ile Perrot. *Le Naturaliste Canadien*, 105, 56–57.
- Razo-Mendivil, U., León-Règagnon, V. & Pérez-Ponce de León, G. (2006) Monophyly and systematic position of *Glypthelmins* (Digenea), based on partial lsrDNA sequences and morphological evidence. *Organisms, Diversity & Evolution*, 6, 308–320.
- Razo-Mendivil, U.J. & Pérez-Ponce de León, G. (2008) Taxonomic revision of the genus *Glypthelmins* Stafford, 1905. *Zootaxa*, 1882, 1–45.
- Reiber, R.J. (1941) Nematodes of Amphibia and Reptilia. I. Reelfoot Lake, Tennessee. *Journal of Tennessee Academy of Sciences*, 16, 92–99.
- Reiber, R.J., Byrd, E.E. & Parker, M.V. (1940) Certain new and already known nematodes from Amphibia and Reptilia. *Lloydia*, 3, 12–144.
- Ridgeway, B.T. (1964) Observations on the morphology and life history of *Oswaldocruzia* sp. in frogs. *Proceedings of the Iowa Academy of Science*, 71, 525–531.
- Rodriguez-Ortiz, B., Garcia-Prieto, L. & Perez-Ponce de Leon, G. (2004) Checklist of the helminth parasites of vertebrates in Costa Rica. *Revista de Biologia Tropical*, 52, 313–354.
- Rosen, R. & Manis, R. (1976) Trematodes of Arkansas amphibians. Journal of Parasitology, 62, 833-834.
- Rudolphi, C.A. (1808) Entoozoorum sive vermium intestinalium historia naturalis. I. XXIV + 527 pp. Amstelaedami.
- Rudolphi, C.A. (1819) Entozoorum synopsis, cui accedunt mantissa duplex et indices locupletimmimi. Augusti Rücker, Berlin, 811 pp.
- Santos-Barrera, G., Hammerson, G., Hedges, B., Joglar, R., Inchaustegui, S., Kuangyang, L., Wenhao, C., Huiqing, G., Haitao, S., Diesmos, A., Iskandar, D., van Dijk, P.P., Matsui, M., Schmidt, B. & Miaud, C. (2006) *Lithobates catesbeianus*. In: IUCN 2008. 2008 IUCN Red List of Threatened Species. <www.iucnredlist.org>. Downloaded on 17 April 2009.
- Schacher, J.F. & Crans, W.J. (1973) *Foleyella flexicauda* sp. n. (Nematoda: Filarioidea) from *Rana catesbeiana* in New Jersey, with a review of the genus and erection of two new subgenera. *Journal of Parasitology*, 59, 685–691.
- Schell, S.C. (1962) Development of the sporocyst generations of *Glypthelmins quieta* (Stafford, 1900) (Trematoda: Plagiorchioidea), a parasite of frogs. *Journal of Parasitology*, 48, 587–593.
- Schell, S.C. (1965) The life history of *Haematoloechus breviplexus* Stafford, 1902 (Trematoda: Haplometridae McMullen, 1937) with emphasis on the development of the sporocyst. *Journal of Parasitology*, 51, 587–593.
- Schrank, F.P. (1788) Verzeichniss der bisher hinlänglich bekannten Eingeweidewürmer, nebst einer Abhandlung über ihre Anverwandtschafen. München, 116 pp.
- Seely, L.B. (1900) Two Distomes. *Biological Bulletin*, 10, 249–254.
- Seno, H. (1907) Distomes of Japan. Zoological Magazine of Tokyo, 19, 121–359.
- Shields, J.D. (1987) Pathology and mortality of the lung fluke *Haematoloechus longiplexus* (Trematoda) in *Rana catesbeiana. Journal of Parasitology*, 73, 1005–1013.
- Shoop, W.L. (1985) Life cycle of Alaria marcianae in Louisiana. Dissertation Abstracts International B, (Sciences and Engineering), 46, 1087–1088.
- Skrjabin, K.I. (1947) Trematodes of Animal and Man. Osnovy Trematodolgii, Moskow, 515 pp.
- Slagle, W.G. (1966) A survey of the helminths parasitic in Rana catesbeiana Shaw and Rana pipiens Schreber from Brazos County, Texas, and vicinity. M.Sc. Thesis, Texas A&M University, College Station, Texas, 45 pp.
- Slom, T.J., Cortese, M.M., Gerber, S.I., Jones, R.C., Holtz, T.H. & Lopez, A.S. (2002) An outbreak of eosinophilic meningitis caused by *Angiostrongylus cantonensis* in travelers returning from the Caribbean. *New England Journal of Medicine*, 346, 668–675.
- Smith, R.J. (1953) *The life history of Megalodiscus ferrissianus n. sp. (Trematoda: Paramphistomatidae)*. Dissertation Ph.D. The University of Michigan.

- Smythe, A.B. & Font, F.F. (2001) Phylogenetic analysis of *Alloglossidium* (Digenea: Macroderoididae) and related genera: life-cycle evolution and taxonomic revision. *Journal of Parasitology*, 87, 386–391.
- Snyder, S.D. & Janovy, J. Jr. (1994) Second intermediate host-specificity of *Haematoloechus complexus* and *Haematoloechus medioplexus* (Digenea: Haematoloechidae). *Journal of Parasitology*, 80, 1052–1055.
- Snyder, S.D. & Tkach, V.V. (2001) Phylogenetic and biogeographical relationships among some holartic frog lung flukes (Digenea: Haematoloechidae). *Journal of Parasitology*, 87, 1433–1440.
- Sprent, J.F.A. (1977) Ascaridoid nematodes of amphibians and reptiles: *Dujardinascaris. Journal of Helminthology*, 51, 253–287.
- Stafford, J. (1900) Some undescribed Trematodes. Zoologischer Jahrbucher Systematic, 13, 399-414.
- Stafford, J. (1902a) Cephalogonimus americanus (new species). Zentralblatt Bakteriologie, 32, 719–725.
- Stafford, J. (1902b) On the representation of *Distomum variegatum*. Zoologische Jahrbuecher Abteilung fuer Systematik Oekologie und Geographie der Tiere, 16, 895–912.
- Stafford, J. (1903) The American representatives of Distomum cygnoides. Zoologiche Jahrbucher, 17, 411–424.
- Stafford, J. (1905) Trematodes from Canadian vertebrates. Zoologische Anzeiger, 28, 681-694.
- Steiner, G. (1924) Some nemas from the alimentary tract of the Carolina tree frog (*Hyla carolinensis* Pennant). *Journal of Parasitology*, 11, 1–32.
- Stunkard, H.W. (1916) On the anatomy and relationships of some North American trematodes. *Journal of Parasitology*, 3, 21–27.
- Stunkard, H.W. (1924) On some trematodes from Florida turtles. *Transactions of the American Microscopical Society*, 43, 97–117.
- Stunkard, H.W. (1973) Observations on *Tubulovesicula pinguis* (Linton, 1910) Manter, 1947 and on the systematics of the hemiurid trematodes. *Biological Bulletin*, 145, 607–626.
- Stunkard, H.W. & Dunihue, F.W. (1933a) Gyrodactylus as parasite of the tadpoles of Rana catesbeiana. Anatomical Record, 57, 98–99.
- Stunkard, H.W. & Dunihue, F.W. (1933b) *Gyrodactylus* as a parasite of the tadpoles of *Rana catesbeiana*. *Journal of Parasitology*, 20, 137.
- Sullivan, J.J. (1972) The status of the "glypthelminth" trematodes with an account of the life-cycle of Hylotrema pennsylvaniensis (Cheng, 1961) n. gen., n. comb. Ph. D. Thesis, University of Georgia, USA 158 pp.
- Sullivan, J.J. (1976) The trematode genus *Glypthelmins* Stafford, 1905 (Plagiorchoidea: Macroderoididae) with a redescription of *G. facioi* from Costa Rican frogs. *Proceedings of the Helminthological Society of Washington*, 43, 116–125.
- Tantaleán, M. & Michaud, C. (2005) Definitive hosts of *Spirometra mansonoides* (Cestoda, Diphyllobothriidae) in Peru. *Revista Peruana de Biología*, 12, 153–157.
- Terwedow, H.A. & Craig, G.B. (1977a) Development of *Waltonella flexicauda*, a filarial parasite of *Rana catesbeiana*, in *Aedes aegypti* and other culicine mosquitoes. *Proceedings of the Helminthological Society of Washington*, 44, 86–91.
- Terwedow, H.A. & Craig, G.B. (1977b) *Waltonella flexicauda*: development controlled by a genetic factor in *Aedes aegypti. Experimental Parasitology*, 41, 272–282.
- Thomas, L.J. (1939) Life cycle of a fluke, *Halipegus eccentricus* n. sp., found in the ears of frogs. *Journal of Parasitology*, 25, 207–221.
- Torres, P. & Puga, S. (1996) Occurrence of cystacanths of *Centrorhynchus* sp. (Acanthocephala: Centrorhynchidae) in toads of the genus *Eupsophus* in Chile. *Memorias do Instituto Oswaldo Cruz*, 91, 717–719.
- Travassos, L. (1921) Contribuções para o conhecimento da fauna helmintolojica brasileira XV. Sobre as species brasileiras da familia Lecithodendriidae Odhner, 1911. Archivos da Escola Superior de Agricultura e Medicina Veterinaria Nichtheroy, 5, 73–79.
- Travassos, L. (1930) Genero Pleurogenoides Travassos, 1921 (Trematoda: Lecithocendriidae). Memorias do Instituto Oswaldo Cruz, 29, 19–178.
- Travassos, L. (1934) Synopse dos Paramphistomidea. Memorias do Instituto Oswaldo Cruz, 29, 19–178.
- Trowbridge, A.H. & Hefley, H.M. (1934) Preliminary studies of the parasite fauna of Oklahoma Anurans. *Proceedings of the Oklahoma Academy of Sciences*, 14, 16–19.
- Ubelaker, J.E. (1965) The taxonomic status of *Langeronia* Caballero and Bravo Hollis, 1949 with the synonymy of *Loxogenes provitellaria*, Sacks, 1952 with *Loxogenes macrocirra* Caballero and Bravo Hollis, 1949. *Transactions of the Kansas Academy of Science*, 68, 187–190.
- Uchida, A. (1975) Checklist of the helminth parasites of Japanese amphibians. *Bulletin Azabu Veterinary College*, 30, 63–81.
- Uchida, A. (1976) Checklist of the helminth parasites of Japanese amphibians (supplement). *Bulletin of the Azabu Veterinary College*, 1, 23–27.
- Uchida, A. & Itagaki, H. (1976) Studies on the amphibian helminthes in Japan. IV. Redescription of *Haematoloechus lobatus* (Seno, 1907) Walton, 1948 (Trematoda, Haematoloechidae) from bullfrogs, *Rana catesbeiana. Japanese Journal of Parasitology*, 25, 360–365.
- Uchida, A. & Itagaki, H. (1980) Distribution of metacercariae of Pharyngostomum cordatum in Aichi Prefecture and path-

ological findings in infected cats. Journal of the Japan Veterinary Medical Association, 33, 594-597.

- Uchida, A., Inoue, H. & Itagaki, H. (1977) Studies on the amphibian helminths in Japan. V. Metacercariae of a feline intestinal fluke *Pharyngostomum cordatum* in amphibian and reptilian hosts from Kagawa Prefecture. *Japanese Journal of Parasitology*, 26, 384–387.
- Ukoli, F.M.A. (1966) On *Clinostomum tilapiae* n. sp., and *C. phalacrocoracis* Dubois, 1931 from Ghana, and a discussion of the systematics of the genus *Clinostomum* Leidy, 1856. *Journal of Helminthology*, 40, 187–214.
- Ulmer, M.J. (1970) Studies on the helminth fauna of Iowa. I. Trematodes of amphibians. *American Midland Naturalist*, 83, 38–64.
- Underwood, H.T. & Dronen, N.O. (1977) The molluscan intermediate hosts for species of *Haematoloechus* Looss 1899 (Digenea: Plagiorchiidae) from raniid frogs of Texas. *Journal of Parasitology*, 63, 122.
- Van Cleave, H.J. & Lynch, J.E. (1950) The circumpolar distribution of *Neoechinorhynchus rutili*, an acanthocephalan parasite of freshwater fishes. *Transactions of the American Microscopical Society*, 69, 156–171.
- Vanderburgh, D.J. & Anderson, R.C. (1987a) The relationship between nematodes of the genus Cosmocercoides Wilkie, 1930 (Nematoda: Cosmocercoidea) in toads (Bufo americanus) and slugs (Deroceros leave). Canadian Journal of Zoology, 65, 1650–1661.
- Vanderburgh, D.J. & Anderson, R.C. (1987b) Preliminary observations on seasonal changes in prevalence and intensity of Cosmocercoides variabilis (Nematoda: Cosmocercoidea) in Bufo americanus. Canadian Journal of Zoology, 65, 1666–1667.
- Vanderburgh, D.J. & Anderson, R.C. (1987c) Seasonal changes in prevalence and intensity of *Cosmocercoides dukae* (Nematoda: Cosmocercoidea) in *Deroceros leave. Canadian Journal of Zoology*, 65, 1662–1665.
- Vulpian, M. (1859) Note sur un noveau distoma de la grenouille (*Distomum ovocaudatum*). Comptes Rendus des Séances et Mémoires de la Société de Biologie, 5, 150–152.
- Waitz, J.A. (1961) Parasites of Idaho amphibians. Journal of Parasitology, 47, 89.
- Waitz, J.A. (1962) Parasitic helminths as aids in studying the distribution of species of *Rana* in Idaho. *Transactions of the Illinois State Academy of Sciences*, 54, 152–156.
- Walters, J.C., Freeman, R.S., Sheas, M. & Fallis, M. (1975) Penetration and survival of mesocercariae (*Alaria* spp.) in the mammalian eye. *Canadian Journal of Ophthalmology*, 10, 101–106.
- Walton, A.C. (1927) A revision of the nematodes of the Leidy collections. *Proceedings of the Academy of Natural Sciences of Philadelphia*, 79, 49–163.
- Walton, A.C. (1929) Studies on some nematodes of North American frogs. Journal of Parasitology, 15, 227–240.
- Walton, A.C. (1931) Note on some larval nematodes found in frogs. Journal of Parasitology, 17, 228-229.
- Walton, A.C. (1933) The nematode as parasites of Amphibia. Journal of Parasitology, 20, 1–32.
- Walton, A.C. (1935) The nematode as parasites of Amphibia. II. Journal of Parasitology, 21, 27-50.
- Walton, A.C. (1937) The Nematoda as parasites of Amphibia. III. Studies on life histories. *Journal of Parasitology*, 23, 299–300.
- Walton, A.C. (1938) The trematodes as parasites of amphibia list of host. *Contribution from the Biological Laboratories of Knox College*, 62, 1–24.
- Walton, A.C. (1940) Notes on amphibian parasites. Proceedings of the Helminthological Society of Washington, 7, 87–91.
- Walton, A.C. (1947) Parasites of the Ranidae (Amphibia). I. Journal of Parasitology, 33 (suplement), 25-26, 26, 26-27.
- Walton, A.C. (1949) Parasites of the Ranidae (Amphibia). VII. *Transactions of the American Microscopical Society*, 68, 49–54.
- Ward, H.B. & Magath, T.B. (1917) Notes on some nematodes from freshwater fishes. Journal of Parasitology, 3, 57-64.
- Whitehouse, C.A. (2002) A study of the frog lung fluke *Haematoloechus* (Trematoda: Haematoloechidae) collected from areas of Kentucky and Indiana. *Proceedings of the Indiana Academy of Sciences*, 1, 67–76.
- Wootton, D.M., Ryan, K.A., Demaree, R.S. & Critchfield, R.L. (1993) A new species of *Gyrodactylus* (Monogenea: Monopisthocotylea) on tadpoles of *Rana catesbeiana* from California, U.S.A. *Transactions of the American Micro-scopical Society*, 112, 230–233.
- Yamaguti, S. (1958) Systema Helminthum. Vol. I, The digenetic trematodes of vertebrates. Part 1. Interscience Publishers, New York, 979 pp.
- Yamaguti, S. (1961) *Systema Helminthum. Vol. III, The nematodes of vertebrates.* Interscience Publishers, New York, Part I, 679 pp.; Part II, 692 pp.
- Yamaguti, S. (1963) Systema Helminthum. Vol.V, Acanthocephala. Interscience Publishers, New York, 423 pp.
- Yamaguti, S. (1971) Synopsis of the digenetic trematodes of vertebrates. Keigaku Publishing Company, Tokyo, 1772 pp.
- Yildirimhan, H.S., Bursey, C.R. & Goldberg, S.R. (2005) Helminth parasites of the Caucasian salamander, *Mertensiella caucasica*, from Turkey. *Comparative Parasitology*, 72, 75–87.
- Yoder, H.R. & Gomez, G.W. (2007) Helminth parasite assemblages in bullfrogs (*Rana catesbeiana*) from southeast Texas. *Texas Journal of Science*, 59, 33–38.
- Zamparo, D. & Brooks, D.R. (2005) Three rarely reported digeneans inhabiting amphibians from Vancouver Island, British Columbia, Canada. *Journal of Parasitology*, 91, 1242–1244.
- Zeder, J.G.H. (1800) Erster Nachtrag zur Naturgeschichte der Eingeweidewürmer mit. Aufüssen und Anmerkungen Her-

ausgegeben, Leipzig, Germany, 320 pp.

- Zelmer, D.A. & Brooks, D.R. (2000) *Halipegus eschi* n. sp. (Digenea: Hemiuridae) in *Rana vaillanti* from Guanacaste Province, Costa Rica. *Journal of Parasitology*, 86, 1114–1117.
- Zelmer, D.A. & Esch, G.W. (1999) Reevaluation of the taxonomic status of *Halipegus occidualis* Stafford, 1905 (Digenea: Hemiuridae). *Journal of Parasitology*, 85, 157–160.

Taxonomic index

Abbreviata 24, 25 Abbreviata ranae 25 Abbreviata sp. 24 Acanthocephala 23 Acanthocephalus ranae 23 Acanthocephalus 23 Acanthocephalus sp. 23 Acuaria 36 Agamascaris odontocephala 25 Agamonema sp. 25 Alaria 3,38 Alaria arisaemoides 3 Alaria marcianae 3 Alaria mustelae 3 Alaria sp. 3 Alaria spp. 4 Allassostomoides chelydrae 4 Allassostomoides parvus 4 Angiostrongylus cantonensis 25 Anisakinae gen. sp. 25 Apharyngostrigea pipientis 4 Aplectana cubana 26 Aplectana haematospicula 26 Aplectana 26 Aplectana sp. 26 Ascarididae gen. sp. 26 Ascaris 26 Ascaris sp. 26 Auridistomum chelydrae 4 Bothriocephalus acheilognathi 21 Bothriocephalus sp. 21 Brachycoelium louisianae 5 Brachycoelium salamandrae 4 Brevimulticaecum sp. 27 Bunodera sp. 5 Camallanus 27 Camallanus multilineatus 27 Camallanus sp. 27 Centrorhynchus 27

Centrorhynchus sp. 23 Cephalogonimus americanus 5 Cephalogonimus brevicirrus 5 Cephalogonimus sp. 5 Cercaria kentuckiensis 17 Cestoidea 21 Clinostomum 38 *Clinostomum attenuatum* 6 Clinostomum complanatum 6 Clinostomum marginatum 6 Clinostomum sp. 5 Contracaecum sp. 27 *Contracaecum* spp. 27 Cosmocercoides 27 Cosmocercoides dukae 28 Cosmocercoides sp. 27 Cosmocercoides variabilis 1, 2, 3, 28 Cylindrotaenia americana 21 Cystagora tetracystis 6 Digenea 3 Dioctophyma renale 28 Dioctophymidae gen. sp. 28 Diphyllobothrium ranarum 23 Diplodiscus 6, 16 Diplodiscus intermedius 16 Diplodiscus sp. 6 Diplodiscus subclavatus 17 Diplodiscus temperatus 17 Distoma cygnoides var. B 10 Distomum arcanum 15 Distomum cygnoides 9, 10 Distomum variegatum 13 Dujardinascaris sp. 29 Dujardinia 29 Echinorhynchus sp. 23 Echinostoma trivolvis 6 Enterobius vermicularis 29 Euryhelmis squamula 7 Eustrongylides 29

Eustrongylides sp. 29 Eustrongylides wenrichi 29 Falcaustra 30 Falcaustra catesbeianae 30 Falcaustra inglisi 30 Falcaustra sp. 29 Fessisentis friedi 23 Filaria nitida 30 Filaria quadrituberculata 30 Foleyella 32 Foleyella americana 31 Foleyella brevicauda brevicauda 31 Foleyella brevicauda magnilarvata 31 Foleyella candezei 31 Foleyella flexicauda 31 Foleyella furcata 31 Foleyella philistinae 31 Foleyella seasonalis 31 Foleyella sp. 31 Foleyellides 31, 32 Foleyellides americana 31 Foleyellides flexicauda 31 Foleyellides ranae 31 Foleyellides sp. 31 Glypthelmins 7,8 Glypthelmins californiensis 7 Glypthelmins hyloreus 7 Glypthelmins intestinalis 7 Glypthelmins linguatula 19 Glypthelmins parva 7 Glypthelmins pennsylvaniensis 7 Glypthelmins proxima 19 Glypthelmins quieta 1, 2, 7, 8, 37 Glypthelmins shastai 7 Glypthelmins sp. 7 Glypthelmins subtropica 8 Gnathostoma spinigerum 32, 38 Gorgodera 9,10 Gorgodera amplicava 8,9 Gorgodera circava 8 Gorgodera cygnoides 9 Gorgodera minima 9 Gorgodera simplex 10 Gorgodera sp. 8 Gorgodera vivata 8 Gorgoderidae gen. sp. 9 Gorgoderina 8 Gorgoderina attenuata 1, 2, 9, 37

Gorgoderina bilobata 10 Gorgoderina simplex 10 Gorgoderina sp. 9 Gyrinicola batrachiensis 32 Gyrodactylus 20 Gyrodactylus arcuatus 20 Gyrodactylus catesbeianae 20 Gyrodactylus jennyae 21 Gyrodactylus sp. 20 Haematoloechus 10, 11, 13, 38 Haematoloechus breviplexus 10, 11, 13 Haematoloechus buttensis 10, 11 Haematoloechus coloradensis 1, 2, 3, 10, 11, 13 Haematoloechus complexus 10, 11, 38 Haematoloechus danbrooksi 12 Haematoloechus floedae 10, 11, 12 13, 14 Haematoloechus lobatus 12 Haematoloechus longiplexus 12 Haematoloechus macrorchis 12 Haematoloechus medioplexus 12, 38 Haematoloechus parviplexus 1, 2, 3, 13 Haematoloechus sp. 10 Haematoloechus variegatus 13 Haematoloechus varioplexus 13 Haematoloechus viguerasi 13 Halipegus 14, 38 Halipegus amherstensis 14 Halipegus eccentricus 14 Halipegus occidualis 14 Halipegus ovocaudatus 15 Halipegus sp. 14 Hedruris 32 Hedruris pendula 32 Hedruris siredonis 32, 33 *Hedruris* sp. 32 Kathlaniidae gen. sp. 33 Langeronia macrocirra 15 Levinseniella ophidea 15 Loxogenes arcanum 15 Loxogenes provitellaria 15 Loxogenes sp. 15 Loxogenes bicolor 16 Loxogenoides 15, 16 Loxogenoides bicolor 16 Loxogenoides loborchis 16 Megalodiscus 16 Megalodiscus americanus 16 Megalodiscus ferrisianus 16

Megalodiscus intermedius 16 Megalodiscus microphagus 7 Megalodiscus sp. 16 Megalodiscus temperatus 16, 17 Mesocoelium brevicaecum 17 Mesostephanus 17 Monogenea 20 Nematoda 24 Neoechinorhynchus 24 Neoechinorhynchus cyanophlytic 24 Neoechinorhynchus rutili 24 Neoechinorhynchus sp. 24 Neogogatea 17 Neogogatea kentuckiensis 17 Onchocercidae gen. sp. 33 Ophiotaenia 21 Ophiotaenia bufonis 21 Ophiotaenia gracilis 21, 22 Ophiotaenia magna 22 Ophiotaenia saphena 22 Ophiotaenia sp. 21 **Opisthoporus** 20 Oswaldocruzia leidyi 33 Oswaldocruzia lenteixeirai 33 Oswaldocruzia pipiens 34 Oswaldocruzia 33 Oswaldocruzia sp. 33 Oxysomatium inglisi 30 Oxysomatium longicaudata 35 Oxysomatium sp. 34 Oxyurida gen. sp. 34 Oxyuris vermicularis 29 Paramphistomidae gen. sp. 18 Paramphistomum aspidonectes 20 Parapharyngodon bassi 34 Pharyngostomum cordatum 18 Phyllodistomum sp. 18 Phyllodistomum 10 Physaloptera ranae 25 Physaloptera sp. 35

Physalopteridae gen. sp. 34 Plagiorchis sp. 18 Pleurogenoides 18 Pleurogenoides sp. 18 Pleurogenoides stromi 18 Pneumobites 10, 11 Pneumonoeces lobatus 12 Porrocaecum sp. 35 Proteocephalidae gen. sp. 22 Proteocephalus sp. 22 Proterometra albacauda 18 Pseudosonsinotrema catesbeianae 19 Raillietnema 35 Raillietnema longicaudata 35 Rauschiella 19 Rauschiella linguatula 19 Rauschiella proxima 19 Rhabdias 35 Rhabdias americanus 35 Rhabdias bufonis 35 Rhabdias ranae 1, 2, 3, 36 Rhabdias sp. 35 Ribeiroia sp. 19 Spinitectus gracilis 1, 2, 36, 37 Spinitectus sp. 36 Spirometra mansonoides 22 Spirometra ranarum 23 Spironoura 30 Spironoura catesbianae 30 Spiroptera 36 Spiroptera mugientis 36 Spiroxys constricta 37 Spiroxys contorta 37 *Spiroxys* sp. 1, 2, 36 Spirura 36 Strigea elegans 19 Strongyluris ranae 37 Teloporia 20 Teloporia aspidonectes 20 Tetrapapillatrema concavocorpa 4